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2 Differential patterns of within- and between-population genetically-based trait variation

- 3 in Lupinus angustifolius L.
- 4 Running title: Within- and between-population traits variation in Lupinus angustifolius L.
- 5 *For correspondence. E-mail cristinapoyatosfernandez@gmail.com
- 6 Authors: Cristina Poyatos^{1,*}, Sandra Sacristán-Bajo¹, Pablo Tabarés¹, Samuel Prieto-Benítez²,
- 7 María Luisa Rubio Teso¹; Elena Torres³, Javier Morente-López^{1,4}, Carlos Lara-Romero¹, José
- 8 María Iriondo¹, Alfredo García Fernández¹
- 9 Affiliations:
- 10 ¹Grupo de Ecología Evolutiva (ECOEVO). Área de Biodiversidad y Conservación,
- Departamento de Biología y Geología, Universidad Rey Juan Carlos-ESCET, Tulipán s/n.
 28933 Móstoles, Madrid (Spain).
- ²Ecotoxicology of Air Pollution, CIEMAT. Avda. Complutense, 40. 28040, Madrid (Spain).
- ³Departamento de Biotecnología-Biología Vegetal, Universidad Politécnica de Madrid, Av.
- 15 Puerta de Hierro 2-4, 28040, Madrid (Spain).
- ⁴Grupo de Ecología y Evolución en Islas, Instituto de Productos Naturales y Agrobiología
- 17 (IPNA-CSIC), Avda. Astrofísico Francisco Sánchez 3, 38206, San Cristóbal de La Laguna,
- 18 Santa Cruz de Tenerife (Spain).
- 19

1 ABSTRACT

Background and aims. Within-population genetic and phenotypic variation play a key role in the development of adaptive responses to environmental change. Between-population variation is also an essential element to assess the evolutionary potential of species in response to changes in environmental conditions. In this context, common garden experiments are a useful tool to separate the genetic and environmental components of phenotypic variation. We aimed to assess within and between-population phenotypic variation of *Lupinus angustifolius* L. in terms of its evolutionary potential to adapt to ongoing climate change.

9 Methods. We evaluated populations' phenotypic variation of foliar, phenological and 10 reproductive traits with a common garden experiment. Patterns of functional trait variation were 11 assessed with 1) mixed models analyses and coefficients of variation (CV) with confidence 12 intervals; 2) principal component analyses (PCA); and 3) correlations between pairs of traits. 13 Analyses were performed at the population level (four populations) and at the latitude level 14 (grouping pairs of populations located in two latitudinal ranges).

15 *Key results.* Phenotypic variation had a significant genetic component associated with a 16 latitudinal pattern. 1) Mixed models found lower SLA, advanced flowering phenology and 17 lower seed production of heavier seeds in southern populations, whereas CV analyses showed 18 lower within-latitude variation especially in phenological and reproductive traits at the southern 19 populations. 2) PCAs showed a clearer differentiation of phenotypic variation between latitudes 20 than between populations. 3) Correlation analyses showed a greater number of significant 21 correlations between traits in southern populations (25 *vs.* 13).

Conclusions. Between-population phenotypic variation was determined by contrasting
 temperature and drought at different latitude and elevation. Southern populations had
 differential trait values compatible with adaptations to high temperatures and drought.

Moreover, they had lower within-population variation and greater number of trait correlations
 probably as a result of these limiting conditions, making them more vulnerable to climate
 change.

4 Key words: *Lupinus angustifolius*, traits, intraspecific variation, population variation,
5 evolutionary potential, latitude, climate change.

6 INTRODUCTION

Phenotype trait expression is the result of a complex set of traits that is determined by the 7 interaction of the genetic component and the environment (Murren, 2012). Climate change 8 9 triggers abrupt environmental changes prompting the need for a trait expression change in a 10 quick adaptive process (Nagahama et al., 2018; Cruz-Maldonado et al., 2021). In this context, 11 many species are threatened because their evolutionary processes take place more slowly than environmental changes due to climate change (Botkin et al., 2007). Therefore, understanding 12 13 the change in phenotypic trait variation along environmental gradients is necessary to develop 14 strategies aimed at the conservation, restoration and management of populations (Mousavi-Derazmahalleh et al., 2018), since species success depends in great part on their intraspecific 15 trait variation (McGill et al., 2006; Pescador et al., 2015; Halbritter et al., 2018; Funk et al., 16 2019). For this reason, the number of studies that have monitored intraspecific trait variation 17 have multiplied in the last few years (e.g., Carvalho et al., 2020; Westerband et al., 2020; Kühn 18 19 et al., 2021; Martin and Isaac, 2021).

Intraspecific trait variation has its basis in individual variation and in the traits' variability
expression. It can be evaluated from the genetic variation and phenotypic plasticity perspectives
(Violle *et al.*, 2012), two qualities that largely determine plants' persistence in a climate change
scenario (Funk *et al.*, 2019; Westerband *et al.*, 2021) by maintaining population fitness in
environments with great climatic variation (Matesanz *et al.*, 2010; Welles and Funk, 2021).

Many studies have used common garden experiments to distinguish the genetic or phenotypic
 plasticity origin of such variation (e.g., Lara-Romero *et al.*, 2014; Moncalvillo *et al.*, 2019;
 Matesanz *et al.*, 2020).

Species' phenotypic variation can be decomposed in two different sources of variability: 4 5 within- and between-populations. Within-population phenotypic variation is an element that determines population success because its genetic component allows evolution by natural 6 selection (Bolnick et al., 2011; Westerband et al., 2021). High within-population adaptive 7 8 genetic variation may favour rapid evolutionary responses that can mitigate climate change effects and limit extinction rates (Sgrò et al., 2011). However, in highly stressful conditions, 9 10 within-population variation may be reduced due to the presence of powerful environmental filters that act above the level of the individual (Pescador et al., 2015; Luo et al., 2016; Helsen 11 12 et al., 2020). These abiotic filters can generate more coordinated phenotypes by increasing the 13 number of correlations between traits (Carvalho et al., 2020). This scenario limits natural selection performance and favours compensation between traits to maintain phenotype viability 14 15 (Milla and Reich, 2011). On the other hand, between-population variation includes differences between populations that can be explained by the evolutionary history of populations, 16 17 comprising isolation or adaptation processes, neutral genetic mechanisms, demographic bottlenecks and biotic and abiotic filters (Ohsawa and Ide, 2008). In this work, we have 18 highlighted the role of latitude and elevation as shapers of abiotic filters. Latitude and elevation 19 model the distribution of many climatic variables generating environmental gradients that may 20 operate at a reduced spatial distance. For instance, increasing latitude is usually associated with 21 22 lower temperatures, greater precipitation, and lower daily solar radiation (Li et al., 1998). Similarly, increasing elevation is usually associated with lower temperatures and greater 23 24 precipitation, but also to higher radiation exposure and snow accumulation (Körner, 2007). In this way, they influence traits' phenotypic expression (Hulshof et al., 2013) and plants' 25

responses, shaping scenarios in which natural selection can play a relevant role in generating
 local adaptations (Ohsawa and Ide, 2008; Halbritter *et al.*, 2018).

Drought and rising temperatures can produce different responses according to the species 3 4 considered. Annual species usually develop a drought escape strategy (Welles and Funk, 2021) 5 that affects traits phenotypic variation. In this way, some foliar traits (e.g. specific leaf area or 6 SLA and leaf dry matter content or LDMC) have been reported to be highly influenced by these climatic conditions (Yulin and Zhang, 2005; Hulshof et al., 2013). In warmer environments, 7 8 leaves tend to reduce evapotranspiration and to promote carbon assimilation (lower SLA and higher LDMC values) (Welles and Funk, 2021). In this sense, they can grow faster to (i) 9 10 advance their phenology (e.g. earlier flowering onset) and (ii) to ensure their seeds contain the necessary assimilates to germinate (e.g. heavier seeds) (Rubio de Casas et al., 2017; Anstett et 11 al., 2021; Welles and Funk, 2021; Matesanz et al., 2022). 12

Here, we evaluated the within- and between-populations phenotypic variation of *Lupinus angustifolius* L. (Fabaceae) regarding foliar, phenological and reproductive traits using a common garden approach. To do so, we used two pairs of *L. angustifolius* populations selected according to their latitudinal divergence and marked climatic/environmental differences (northern *vs.* southern populations). The main objective of this study was to assess the genetically based phenotypic variation of *L. angustifolius* populations in terms of their evolutionary potential to respond to environmental threats derived from climate change.

20 In this context, we addressed the following hypotheses:

We consider that between-population phenotypic variation will mostly originate from
 divergent selection processes resulting from the contrasting temperature and water availability
 conditions caused by differences in latitude and elevation. Therefore, we expect between population phenotypic variation to be greater at the latitudinal level (northern *vs.* southern

populations) than at the within-latitude level (differences between populations at a given
 latitude) and that the trait values obtained at each location will be associated with adaptations
 to high temperature and water availability constraints.

2) Southern latitude populations of *L. angustifolius*, occurring at higher temperatures and
aridity, will experience more intense selective pressures (i.e., drought tolerance, early flowering
onset, etc.). We expect southern populations to have a greater number of significant correlations
between traits and less variation within and between populations compared to northern
populations, since the intensity of selective pressures has been positively associated with
phenotypic integration (i.e., the correlation of multiple functionally related traits) (Pigliucci,
2003; Carvalho *et al.*, 2020).

3) Because higher selective pressures concomitant to southern populations are associated with 11 12 specific limitations (i.e., high temperature, drought), the effect of these pressures will be more 13 marked in functional traits more closely associated to these limitations. Therefore, we expect 14 that between-population phenotypic variation at the contrasting latitudes will differ depending on the group of traits analysed (foliar, phenological and reproductive). Specifically, we 15 anticipate that warmer environment populations will reduce water loss by lowering SLA and 16 17 increasing LDMC values. In addition, we expect that they will advance the flowering phenology and produce larger seeds to guarantee their viability as part of an escape strategy to drought. 18

19 MATERIAL AND METHODS

20 Species and study site

Lupinus angustifolius is an annual self-pollinated herbaceous plant distributed across the
 Mediterranean Basin, occurring preferably in well-drained sandy acid soils, including disturbed
 or abandoned areas (Castroviejo and Pascual, 1999; Talhinhas *et al.*, 2006). This legume has

palmate leaves with narrow leaflets and its racemose inflorescence contains bluish-purple 1 flowers. Under natural conditions, L. angustifolius flowers tend almost exclusively to self-2 pollinate, as self-pollination takes place in early stages of development when the flower is still 3 closed (Kazimierska and Kazimierski, 2002). Its legume fruits have variable seed content, 4 commonly between three and six (Castroviejo and Pascual, 1999). The seed presents a hard 5 cover that allows it to remain dormant until environmental conditions are adequate (Berger et 6 7 al., 2017). The flowering season takes place between March and August (Castroviejo and Pascual, 1999). 8

The western half of the Iberian Peninsula is considered the genetic diversity centre of the 9 10 species and it is where this species is most widely distributed (Mousavi-Derazmahalleh et al., 2018). In 2016, two pairs of L. angustifolius populations occurring in the Iberian Peninsula 11 12 were selected according to their latitudinal divergence, showing marked differences in 13 temperature and precipitation (Table 1). Two of them, Zafrón (hereafter, FRO) and Zarapicos (hereafter, PIC) are located in Central Spain in Salamanca province, whereas the other two, La 14 15 Garranchosa (hereafter, GAR) and Rivera de la Lanchita (hereafter, RIV) are located in southwestern Spain in Badajoz province. The latter two populations are located at lower latitude 16 17 and elevation than the former. They also experience higher temperatures and lower water availability (Table 1). The populations were selected over others that were visited because they 18 19 all had hundreds of individuals suggesting more stable population dynamics. In each population, 100 mother plants were haphazardly selected and their seeds were collected 20 21 separately. The experiment developed in the present work used individuals descended from 22 those originally sampled in 2016, since this work is part of a more extensive project (EVA project, AdAptA-lab, Rey Juan Carlos University, Móstoles, Madrid, http://adapta-23 24 lab.com/eva/).

Comentado [1]: ¿Por qué lo pusimos con mayúscula? ¿en inglés se pone así o lo corregimos y ponemos en minúscula?

7

25 Common garden experiment

A common garden experiment was established in randomised blocks at the Rey Juan Carlos 1 University (Móstoles, Madrid, CULTIVE laboratory https://urjc-cultive.webnode.es/; Table 1) 2 to minimise the effect of environmental conditions and assess the genetic component in 3 phenotypic trait expression. In November 2016 three seeds per each sampled mother plant of 4 the species were subjected to manual mechanical scarification with a nail cutter, sown in a six-5 litre (L) pot with a 1:1 mix of commercial substrate and sand, and placed in a greenhouse 6 7 irrigated by nebulisation. After germination and seedling emergence, the largest seedling was selected, and the other two were clipped. In March 2017 L. angustifolius individuals were 8 moved to the acclimation area outside the greenhouse and grown with drip irrigation until plant 9 senescence (June 2017). For two additional seasons (2018 and 2019), a new generation of L. 10 11 angustifolius was grown in the common garden experiment using seeds obtained by natural self-pollination. Each season followed the same schedule: sowing in November, moving to 12 outside facilities in March and plant senescence in June. After the first generation 25% of 13 genotypes in each population were randomly selected and maintained with three siblings per 14 15 genotype. When we mention "season", we are alluding to the period between November and June, that is, from the time we started working with the seeds until we collected the fruits. 16

17 Trait measurements

Twelve functional traits were measured for L. angustifolius individuals during the 2018-2019 18 season (Supplementary data Table S1) to avoid the variation associated with maternal effects 19 in the first two seasons (2016-2017 and 2017/2018). In November 2018, seeds were weighed 20 prior to sowing, and, once sown, the pots were monitored daily in November and December 21 2018 to record the date of emergence of the individuals and obtain seed germination time. In 22 23 March 2019, we recorded the date when the first flower began to bloom for each plant. Thus, 24 we defined flowering onset as the number of days from the date of sowing to the date of 25 appearance of the first flower. The recording of flowering end dates and leaf collection to

measure foliar traits began in April 2019. Finally, the fruits produced by each plant were
 collected in May 2019 to count the number of seeds per fruit.

3 - Foliar traits

Six foliar traits were obtained by performing calculations and direct measurements on leaves: 4 5 1) sum of leaf area (FA), 2) specific leaf area (SLA), 3) relative water content (RWC), 4) leaf dry matter content (LDMC), 5) nitrogen content (N) and 6) phosphorus content (P). As the 6 7 Lupinus angustifolius leaf is palmately compound (with five leaflets that are attached in a single 8 point at the end of the petiole), these traits were measured by collecting the central leaflet of eight randomly chosen leaves per individual. FA was measured by scanning the leaflets 9 (CanoScan LiDE 210; Canon; Japan, Tokio) and adding the leaflet areas obtained with ImageJ 10 software (Abramoff et al., 2004). SLA resulted from the ratio of FA and dry mass (measured 11 12 in grams as the sum of the dry mass of the eight leaflets). The leaflets were dried in an oven at 60°C for at least 72 hours (Digiheat-TFT; SELECTA; Spain, La Rioja). RWC was obtained as 13 14 a percentage resulting from the following formula: [(Mf-Md)/(Ms-Md)] x 100, where Mf, Md and Ms are the fresh mass, dry mass and saturated mass of the leaflets sampled measured in 15 grams (g), respectively. Dry mass was measured with an XPR microbalance (Mettler-Toledo; 16 Greifensee, Switzerland), whereas fresh and saturated mass were measured with an analytical 17 balance of KERN ABJ-ABS series (Solent Scale; Chichester, United Kingdom). LDMC is the 18 percentage obtained from the ratio of dry and fresh mass of the leaflets. Finally, we performed 19 20 the digestion of 50 milligrams (mg) of leaf sample according to the Kjeldahl method (Radojevic 21 and Bashkin, 2015). Subsequently, Skalar protocols were used to obtain total nitrogen and phosphorus content (Skalar San++ autoanalyzer; Skalar, Netherlands). These analyses were 22 23 performed at the Rey Juan Carlos University (Móstoles, Madrid, NUTRILAB laboratory 24 https://nutrilab-urjc.es).

1 - Phenological traits

Four phenological traits were recorded: 1) seed germination time (Germination time), 2) flowering onset (Onset), 3) flowering end (End), and 4) flowering duration (Duration). Germination time was defined as the days elapsed from seed sowing to seed emergence. Onset and End were calculated as the days elapsed from the date of seed emergence to the appearance of the first open flower (Matesanz *et al.*, 2020), and the date the plant stopped producing new flowers, respectively. Duration was calculated as the days elapsed from flowering onset to flowering end.

9 - Reproductive traits

Two reproductive traits were measured: 1) germinated seed weight (Seed mass) and 2) average number of seeds per fruit (Seeds per fruit). Each seed mass was individually weighed in mg using an XPR microbalance (Mettler-Toledo; Greifensee, Switzerland). Seeds per fruit was obtained from 15 fruits randomly obtained from each individual (Supplementary data Table S1). These traits were measured as they are known to influence *L. angustifolius* reproductive success in water stressed environments (Helsen *et al.*, 2020).

16 Statistical analyses

- All statistical analyses were performed using the R statistical environment (version 4.0.3) (R
 Core Team, 2020). To analyse the data at different spatial levels (population and latitudinal
 region), *L. angustifolius* populations were classified according to their latitude as "north" (FRO
 and PIC) or "south" (GAR and RIV) (Table 1).
- First, trait responses were analyzed using Linear Mixed Models (LMM) and Generalized Mixed
 Models (GLMM) at both spatial levels (population and latitudinal region). For this approach,
 we use the glmer and lmer functions within the *lme4* package version 1.1-31 (Bates *et al.*, 2022).
- 24 We defined LMM for continuous response variables (i.e., continuous traits: FA, SLA, RWC,

LDMC, N, P, Seed mass and Seeds per fruit) and GLMM for discrete response variables (i.e., 1 discrete traits: Germination time, Onset, End and Duration). We used a Gaussian distribution 2 for continuous traits and a Poisson distribution for discrete traits. In all models, mother and 3 block variables were introduced as random variables to control the variance due to maternal 4 effects and the variance associated with position within the common garden experiment. At the 5 population level, we included the variable population as a fixed variable and, at the latitudinal 6 7 level, we nested the variable population inside the fixed variable latitude. The significance of fixed variables was determined by a type-II Wald chi-square statistic test. The significance test 8 9 was obtained from the anova function within the car package version 3.1-1 (Fox and Weisberg, 10 2019). To study the existence of significant differences in trait values between population pairs, 11 we then performed a post hoc analysis using the Bonferroni adjustment with pairwise.t.test function in *pairwiseCI* package version 0.1-27 (Schaarschmidt and Gerhard, 2019). 12

Within and between population variation of each trait was estimated by calculating its coefficient of variation (CV). The 95% confidence intervals of the coefficients of variation were calculated using the cv_versatile function of the R package *cvcqv* version 1.0.0 (Beigy, 2019). Comparison of CV confidence intervals between populations was performed using the inference described by Cumming (2009). Then, CV was represented by violin plots applying the vioplot function in *vioplot* R package version 0.3.7 (Adler *et al.*, 2020).

To visualize patterns of similarity between sample units at the population and latitude levels we performed principal component analyses (PCA) (Abdi and Williams, 2010) using the PCA function of the *FactoMineR* package version 2.4 (Lê *et al.*, 2008) which automatically standardise the variables. Leaf nitrogen and phosphorus content were not included in this analysis due to insufficient sample size. PCA results were represented with the *factoextra* package version 1.0.3 (Kassambara and Mundt, 2016).

Finally, Spearman correlation matrices were obtained with the R package Hmisc version 4.6-0 1 (Harrell Jr and Dupont, 2020) for all pairs of traits at the different study levels, and then 2 represented using R packages igraph version 1.2.7 (Csardi and Nepusz, 2006) and ggraph 3 version 2.0.5 (Pedersen, 2021). In addition, correlation matrices between populations or 4 between latitudes were compared using the cortest.mat function of the psych R package version 5 2.1.9 (Revelle, 2021) which develops a chi-square (χ^2) test considering the differences between 6 7 all possible pairs of correlations. This test is generated against the null hypothesis of no differences between the correlation matrices (Carvalho et al., 2020). 8

9 RESULTS

10 Within- and between-population variation and latitude differentiation

11 Mean values differed in a greater number of traits at the population level (nine) than at the latitudinal level (seven) (Table 2). Both levels showed significant differences for FA, SLA, 12 13 Seed mass, Onset, End and Duration (Table 2), but population level also showed significant 14 differences for RWC, LDMC and Seeds per fruit (Table 2). Models detailed information is provided in Supplementary data (Tables S3, S4, S5 and S6). Differences between populations 15 were greater between populations at different latitudes than between populations at the same 16 latitude (Supplementary data Table S7). In particular, RIV was the southern population with 17 the greatest number of significant differences in trait means with northern latitude populations 18 19 (Supplementary data Table S7). Between populations of the same latitude, FRO and PIC populations significatly differed in FA, LDMC and Seeds per fruit, whereas GAR and RIV 20 populations differed in SLA, RWC, LDMC, Onset, Duration and Seed mass traits 21 (Supplementary data Table S7 and Figs. S1 and S2). 22

The first two principal components of the PCAs explained 48.40% of the variation in *L. angustifolius* (Fig. 1 A). PC1 axis had an important contribution from Onset, End and Seed

mass, whereas, in the PC2 axis, LDMC, Germination time and SLA were important traits (Fig. 1 1 A; Supplementary data Table S8). In general terms, L. angustifolius individuals had similar 2 traits according to their population of origin (Fig. 1 B). This is supported by the significant 3 differences obtained at the population level for most of the studied traits (Table 2). Within-4 population phenotypic variation differed depending on the studied trait (Table 3). Thus, the 5 phenotypic variation of five of the study traits was higher in FRO and PIC populations, whereas 6 7 in four other traits, GAR was the population with the greatest within-population variation. Finally, population RIV had the least within-population variation in most of the traits (Table 3, 8 Fig. 1 B). 9

10 From a latitudinal perspective, southern individuals were more clustered to each other than northern ones (Fig. 1 C). When we grouped L. angustifolius populations in northern and 11 12 southern latitudes, we found fewer significant differences between the mean values of all traits, i.e., nine traits with significant differences at the population level versus seven at the latitude 13 level (Table 2). Thus, northern latitude populations showed larger FA, finer leaves (higher SLA) 14 and higher P (Table 2; Supplementary data Table S2 and Fig. S3). The CV of FA was 15 significantly higher at the northern latitude, while RWC showed a significantly higher CV at 16 17 the southern latitude (Supplementary data Table S7).

Flowering onset and end dates were later in the northern latitude populations, and duration was shorter (FRO and PIC, Tables 2; Supplementary data Table S2 and Fig. S4). Onset, End and Duration showed significantly greater within-latitude variation in the northern latitude populations, while Germination time showed greater variation at the southern latitude (Supplementary data Table S2). Seeds from the northern latitude populations had lower mass. Variation in Seed mass and Seeds per fruit was also significantly greater in the northern latitude populations (Supplementary data Table S2 and Fig. S4).

1 Correlation patterns between traits

All trait correlation matrices were significantly different from one another, both at the 2 population and latitudinal levels. A more complex correlation network was observed in GAR 3 4 and RIV (with 15 and 23 significant correlations, respectively) than in FRO and PIC (with 11 5 and 13 significant correlations, respectively) (Fig. 2A, Supplementary data Tables S9 and S10). 6 FRO, GAR and RIV populations presented significant correlations between the three trait blocks (foliar, phenological, and reproductive), whereas PIC presented significant correlations 7 8 only between phenological and foliar traits (Fig. 2A, Supplementary data Tables S9 and S10). The southern and northern latitudes presented 25 and 13 significant trait correlations, 9 10 respectively, nine of which were common to both latitudes (Fig. 2B, Supplementary data Table S11). Additionally, the number of significant correlations between reproductive traits was 11 12 greater in the southern latitude populations (five in the southern latitude versus one in the northern latitude) (Fig. 2B, Supplementary data Table S11). 13

14 DISCUSSION

15 Common garden experiments allow the evaluation of phenotypic traits of plants from different origins under a common and homogeneous environment, reducing to negligible the effect of 16 the environmental component in the expression of the phenotypes (de Villemereuil et al., 2016). 17 Therefore, the differences observed in the phenotypic values can be attributed to genetic 18 19 variation. This way, our common garden experiment provided an assessment of the genetic basis of the within- and between-population phenotypic variation in the studied Lupinus 20 angustifolius populations, as well as some insight on their evolutionary potential to respond to 21 climate change (Lara-Romero et al., 2014). The results of the PCAs (Fig. 1 B and C) in 22 combination with the CVs values at both study levels (Table 3, Supplementary data Table S2) 23 24 showed a larger phenotypic variation when the populations from different latitudes were

compared. Futhermore, several of the trait values obtained at each location (Table 2 and 3, 1 Supplementary data Table S2) can be associated with adaptations to high temperature and water 2 availability constraints, supporting, thus, our first hypothesis. The results obtained also 3 supported our second hypothesis with lower phenotypic variation in most traits in the southern 4 latitude populations (Supplementary data Table S2) and greater number of significant 5 correlations between traits (Supplementary data Tables S9, S10 and S11). Finally, as predicted 6 7 in our third hypothesis, between-population phenotypic variation differed depending on the group of traits analysed. We expected a more marked differential pattern on foliar traits as they 8 are known to be closely related to the drought and high temperature environmental filters that 9 10 mainly affect the southern populations. However, differences in phenological traits were, in this 11 case, also very determinant allowing southern populations to develop an escape strategy to 12 drought.

13 Greater phenotypic variation at the latitudinal level

14 The distribution of individuals in the PCAs allowed us to identify a greater dispersion of phenotypes at the latitudinal level (Fig. 1B and 1C). This is supported by the knowledge that 15 environmental filters at local levels (population level) favor similar phenotypes (Gaüzère et al., 16 2022), i.e., lead to a reduction of phenotypic variation. Combining this observation with the 17 CVs detected at both levels (population and latitudinal), allowed us to conclude that there is 18 greater phenotypic variation at the latitudinal level than between populations (Tables 3, 19 Supplementary data Table S2). For example, the clustering of RIV and GAR slightly diluted 20 the lower CV values of the RIV population for some of the traits (e.g., FA, SLA, RWC, LDMC, 21 N, P, Germination time, Duration, Seed mass and Seeds per fruit). As we predicted, phenotypic 22 23 variation was greater on the latitudinal level for some of the traits. Thus, this pattern of phenotypic variation at the latitudinal level indicated that latitude acts as a potential 24 25 environmental filter (Hulshof et al., 2013). As Ohsawa and Ide (2007) suggest, this may be due

1 to the greater environmental differences existing at this scale, favouring more contrasted

2 between-population variation.

3 Greater number of significant correlations between traits and less variation within and

4 between populations in southern populations

5 Southern populations were more limited in their phenotypic variation probably due to the presence of more restrictive abiotic filters (e.g., more intense drought and aridity), supporting 6 7 the observations of Anderegg et al. (2020) and Carvalho et al. (2020). We noted this in the 8 lower phenotypic variation found in many traits (lower CV) at the southern latitude (e.g., FA, N, Onset, End, Duration, Seed mass and Seeds per fruit) (Supplementary data Table S2). Thus, 9 higher temperatures and lower water availability generated more challenging environmental 10 conditions reducing the number of available phenotypes (and, thus, genotypes) (Sgrò et al., 11 12 2011; Pescador et al., 2015; Luo et al., 2016; Carvalho et al., 2020; Helsen et al., 2020) in L. angustifolius southern populations. 13

The increase in the number of correlations between traits found in *L. angustifolius* southern populations (Fig. 2), in addition to lower phenotypic variation, presents a more complicated situation for these populations. The combination of these two elements generates more coordinated phenotypes, resulting in the process of phenotypic integration (Carvalho *et al.*, 2020). The occurrence of these trade-offs may discard phenotypes that could favour the survival of *L. angustifolius* southern populations under changing environmental conditions. This ultimately could put populations at risk (Anstett *et al.*, 2021).

For future studies, it would be interesting to add other species to integrate characteristics of the
species that are known to affect phenotypic variation (e.g., life cycle and reproductive strategy).
In this sense, *L. angustifolius* is an annual species with an autogamous reproductive strategy
(Castroviejo and Pascual, 1999). An annual strategy triggers a higher number of correlations

and constrictions between traits (Carvalho *et al.*, 2020) by undergoing faster evolution (with
 greater loss of genetic variation) (Anstett *et al.*, 2021). The autogamous character contributes
 to reducing population genetic variation, favouring population isolation and local adaptation
 events.

5 Phenotypic variation varies according to the group of traits studied

6 Our third hypothesis was supported by the differences found in phenotypic variation according 7 to the trait groups formed (foliar, phenological and reproductive). This underlines the 8 importance of studying a variety of traits when assessing the evolutionary potential of a species 9 and, therefore, its capacity to respond to various threats (Westerband *et al.*, 2021).

10 - Foliar traits

The lower SLA values (a consequence of lower FA) obtained in the southern latitude 11 12 populations (Supplementary data Table S2) could be a strategy aimed at reducing leaf evapotranspiration in warmer and drier environments (Pescador et al., 2015; Matesanz et al., 13 2020). Thus, water constraints seem to play a fundamental role in some foliar traits at southern 14 latitudes (Pescador et al., 2015). In northern latitudes the response is likely to be different. 15 16 Summer drought is less intense in northern latitudes of L. angustifolius (FRO and PIC) where 17 annual temperatures are lower (Table 1). This allows L. angustifolius individuals to extend their capture of resources (higher SLA) and develop a strategy of rapid biomass production (Yulin 18 19 and Zhang, 2005). At the same time, lower LDMC values would be expected in northern L. angustifolius populations since SLA and LDMC are known to be negatively correlated. 20 21 However, the relationship between SLA and LDMC weakens when nutrient limitation becomes the main determining factor (Yulin and Zhang, 2005). 22

23 - Phenological traits

Phenological trait responses to an environmental change are essential to ensure population 1 viability (Nagahama et al., 2018). When such responses are triggered in the flowering period, 2 their reproductive success may be affected (Nagahama et al., 2018; Buonaiuto and Wolkovich, 3 2020). Flowering period is a complex process that depends on multiple factors defined by the 4 environment itself. These could be summarized as (i) cold temperatures from autumn to late 5 winter, (ii) warm temperatures from late winter to early spring, and (iii) photoperiod (Ettinger 6 7 et al., 2020). In this study, flowering phenology was much earlier in southern latitude populations of L. angustifolius (GAR and RIV) than in northern populations (FRO and PIC) 8 9 (Table 3). We infer that southern populations of L. angustifolius have developed a drought 10 escape strategy by expressing an earlier flowering onset under environmental conditions that 11 do not allow them to extend their life cycle (Anstett et al., 2021; Welles and Funk, 2021; Matesanz et al., 2022). For this reason, within the same project, a field experiment has been 12 developed for verifying the results obtained in this section (Sacristán-Bajo, under preparation). 13 With this experiment, we could specify that this pattern favours GAR and RIV reproductive 14 15 success in a warmer environment with lower water availability (Matesanz et al., 2020). Southern latitude populations of L. angustifolius showed greater variation in germination time 16 17 according to their CVs. This provides a greater response capacity to adjust to different and 18 unpredictable environmental conditions (Sgrò et al., 2011). Thus, seeds can remain dormant to germinate when conditions become favourable (Rubio de Casas et al., 2017). 19

20 - Reproductive traits

L. angustifolius reproductive traits showed marked patterns between latitudes. Southern
populations GAR and RIV had lower seed production, but the seeds were heavier. This could
be interpreted as an adaptation to drought (Rubio de Casas *et al.*, 2017; Matesanz *et al.*, 2020).
Greater seed mass is related to a strategy focused on ensuring individual survival until it can
reach reproductive maturity in stressful environments (Helsen *et al.*, 2020). However, this

reduces the growing season and the maturation time to seed and fruit (Primack, 1987). Reducing
 seed maturation time triggers a trade-off, reducing the number of seeds produced (Helsen *et al.*,
 2020).

4 CONCLUSION

5 The genetic basis of phenotypic variation studied in the present work allows us to assess the potential of Lupinus angustifolius to generate an adaptive response to climate change. 6 7 Moreover, the evaluation of within- and between-population variation in different functional 8 traits was highly relevant for identifying the different environmental conditions that shape variation in the phenotypic traits of populations and the potential of populations to deal with 9 environmental changes. In this context, populations with lower within-population variation and 10 more coordinated phenotypes (GAR and RIV) may be more vulnerable to swift environmental 11 12 changes because it reduces their ability to adapt. The assessment of different groups of functional traits showed that phenological traits were the most determinant for phenotypic 13 14 expression in L. angustifolius. The reproductive system and life cycle of Lupinus angustifolius conditioned the traits pattern, as well as the latitudinal and altitudinal differences of the 15 populations studied. Finally, this work highlights the role of the study of within- and between-16 17 populations traits variation in predicting populations' ability to face climate change.

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- Fig. 1. Principal component analyses of *Lupinus angustifolius*. The x axis (PC1) represents principal component one, while the y axis (PC2) the principal component two. Variable contributions in the construction of the principal components (A1 and A2) are represented with a colour scale from orange (high contribution) to cyan (low contribution). In each axis (A1 and

A2), percentage of variance explained by each principal component is indicated. Principal
 component analyses at the population level (B1 and B2) and latitudinal level (C1 and C2)
 represent individuals by points with different colors and shapes depending on the population
 (B1 and B2) or latitude (C1 and C2) to which they belong.

Fig. 2. Network of significant correlations of Lupinus angustifolius populations (A) and 5 6 latitudes (B). Arrows symbolize the type of significant correlation obtained: continuous line = positive correlation; dashed line = negative correlation. The size of the arrows indicates the 7 8 value of the correlation: correlation values between 0/0.29 and 0/-0.29, 0.5-point size; values between 0.3/0.49 and -0.3/-0.49, 1-point size; and values 0.5 and above and equal or below -9 10 0.5, 1.5-point size. Foliar traits are represented by green: sum of leaf area (FA), relative water content (RWC), specific leaf area (SLA), leaf dry matter content (LDMC), phosphorus (P) and 11 12 nitrogen content (N); phenological traits by pink: seed germination time (Germination time), flowering onset, end and duration (Onset, End and Duration); and reproductive traits by blue: 13 germinated seed weight (Seed mass) and average seeds per fruit (Seeds per fruit). 14

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Table 1. Geographic coordinates and climatic variables corresponding to the studied populations of *Lupinus angustifolius*. The climatic variables are represented as mean annual temperature (T) measured in degrees Celsius, and May-July precipitation (P) measured in millimeters; elevation is measured in meters. Latitude to which each population belongs (north

or south) is also denoted. Climatic data of *L. angustifolius* populations belong to the 1985-2015
 time series and common garden data located in Móstoles (Madrid) belong to the 2017-2020
 time series. Data were obtained from Sacristán-Bajo *et al.*, (2023).

Localities (population)	Latitu de	Geographic coordinates	Elevation	Т	Р
Zafrón (FRO)	North	41.024192N; 6.028155W	840	12.4	92
Zarapicos (PIC)	North	41.004358N; 5.813066W	820	12.6	89
Rivera de la Lanchita (RIV)	South	38.351586N; 6.576084W	352	16.8	61
La Garranchosa (GAR)	South	38.325735N; 6.433735W	422	16.5	64
Common garden (Mós	toles) 2017			15.1	72
Common garden (Mós	toles) 2018	40.334615N; 3.882168W	690	14.6	95
Common garden (Mós	toles) 2019			14.8	16
Common garden (Mós	toles) 2020			15.1	69
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10 Table 2. Chi-square statistic (X^2) , degrees of freedom (Df) and p-value of type II Wald chi

11 squared tests for all GLMM and LMM generated on population and latitude level. Significant

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values are indicated by the p-value when <0.05 = *, <0.01 = ** and <0.0001 = ***.
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Fixed effects	X^2	D f	p-value	Fixed effects	X^2	Df	p-value
FA				Seed mass			

Population	57.3 0	3	***	Population	296	3	***
Latitude	4.98	1	*	Latitude	98.52	1	***
SLA				Seeds per fruit			
Population	75.3 9	3	***	Population	168.5 2	3	***
Latitude	4.97	1	*	Latitude	2.23	1	0.14
RWC				Onset			
Population	12.7 1	3	**	Population	173.7 1	3	***
Latitude	1.30	1	0.25	Latitude	73.94	1	***
LDMC				End			
Population	57.2 5	3	***	Population	34.53	3	***
Latitude	3.22	1	0.07	Latitude	33.62	1	***
Ν				Duration			
Population	0.72	3	0.87	Population	102.5 3	3	***
Latitude	0.05	1	0.83	Latitude	8.99	1	**
Р				Germination time			
Population	7.83	3	0.05	Population	3.38	3	0.34
Latitude	3.99	1	*	Latitude	2.19	1	0.14

Table 3. Mean, standard deviation (SD) and coefficients of variation (CV) with 95% confidence
intervals for leaf traits: 1) sum of leaf area (FA), 2) specific leaf area (SLA), 3) relative water
content (RWC), 4) leaf dry matter content (LDMC), 5) nitrogen (N) and 6) phosphorus (P)
content; phenological traits: 1) seed germination time (Germination time), 2) flowering onset
(Onset), 3) end (End) and 4) duration (Duration); and reproductive traits: 1) weight of

1 germinated seed (Seed mass) and 2) average of seeds per fruit (Seeds per fruit), from Lupinus

angustifolius populations (FRO, PIC, GAR and RIV).

		Mean ±	SD		CV			
	FRO	PIC	GAR	RIV	FRO	PIC	GAR	RIV
FA (cm ²)	$\begin{array}{c} 14.39 \pm \\ 3.10 \end{array}$	16.81 ± 4.79	$\begin{array}{c} 13.15 \pm \\ 2.61 \end{array}$	11.93 ± 2.24	21.47 (17.57; 26.42)	28.42 (23; 33.95)	19.79 (17.10; 23.05)	18.69 (16.13; 21.52)
SLA (cm ² /g)	211.31 ±32.78	$\begin{array}{c} 216.42 \pm \\ 31.63 \end{array}$	198.30 ± 30.17	$\begin{array}{c} 174.27 \\ \pm 27.08 \end{array}$	15.46 (12.85; 18.33)	14.56 (12.29; 17.25)	15.17 (12.49; 18.30)	15.49 (13.14; 18.21)
RWC (%)	84.91 ± 4.29	83.82 ± 3.92	$\begin{array}{c} 84.66 \pm \\ 5.98 \end{array}$	$\begin{array}{c} 86.80 \pm \\ 4.31 \end{array}$	5.04 (4.28; 5.87)	4.66 (3.83; 5.60)	7.04 (5.87; 8.42)	4.95 (4.02; 6.04)
LDMC (%)	$\begin{array}{c} 15.28 \pm \\ 1.07 \end{array}$	14.69 ± 1.11	13.76 ± 1.25	$\begin{array}{c} 14.57 \pm \\ 0.92 \end{array}$	6.99 (5.94; 8.22)	7.49 (5.62; 9.46)	9.03 (7.25; 10.81)	6.30 (5.16; 7.47)
N (mg/g)	50.61 ± 13.67	50.71 ± 13.48	$\begin{array}{c} 52.85 \pm \\ 14.64 \end{array}$	$\begin{array}{c} 49.70 \pm \\ 10.71 \end{array}$	26.76 (21.50; 33.86)	26.32 (17.12; 36.38)	36.69 (17.48; 54.20)	21.33 (14.80; 29.79)
P (mg/g)	$\begin{array}{c} 4.88 \pm \\ 1.16 \end{array}$	4.57 ± 0.96	$\begin{array}{c} 4.60 \pm \\ 1.05 \end{array}$	$\begin{array}{c} 3.81 \pm \\ 0.78 \end{array}$	23.52 (16.50; 32.03)	20.69 (12.39; 30.48)	33.01 (11.71; 49.61)	20.17 (15.93; 25.79)
Germination time (days)	9 ± 2	9 ± 2	9 ± 3	9 ± 2	22 (17; 27)	20 (17; 23)	28 (23; 34)	20 (17; 24)
Onset (days)	116 ± 8	118 ± 10	103 ± 5	99 ± 6	7 (5; 9)	8 (7; 10)	5 (4; 6)	6 (5; 7)
End (days)	165 ± 6	165 ± 6	155 ± 4	157 ± 5	4 (3; 5)	4 (3; 6)	3 (2; 3)	3 (3; 4)
Duration (days)	47 ± 9	47 ± 8	51 ± 5	57 ± 6	19 (14; 23)	17 (14; 20)	10 (8; 13)	10 (7; 12)
Seed mass (mg)	112.60 ± 18.25	$\begin{array}{c} 106.56 \pm \\ 21.32 \end{array}$	$\begin{array}{c} 163.83 \\ \pm \ 23.48 \end{array}$	154.99 ± 17.95	16.15 (13.24; 19.61)	19.94 (15.93; 24.35)	14.29 (12.64; 15.98)	11.55 (9.48; 13.49)
Seeds per fruit	$\begin{array}{c} 5.75 \pm \\ 0.32 \end{array}$	5.19 ± 0.42	5 ± 0.32	5.07 ± 0.29	5.47 (4.42; 6.63)	7.97 (6.49; 9.63)	6.39 (4.70; 8.07)	5.79 (4.47; 7.19)