

1                   **Evolutionary history of the buildup and breakdown of**  
2                   **the heterostylous syndrome in Plumbaginaceae**

3  
4                   Joana Costa<sup>1\*</sup>, Rubén Torices<sup>2</sup>, Spencer C. H. Barrett<sup>3</sup>

5  
6                   <sup>1</sup> *Centre for Functional Ecology, Department of Life Sciences, University of Coimbra,*  
7                   *Calçada Martim de Freitas, 3000-456 Coimbra, Portugal*

8                   <sup>2</sup> *Area of Biodiversity and Conservation, Universidad Rey Juan Carlos*  
9                   *c/ Tulipán s/n., E-28933. Móstoles, Madrid, Spain*

10                  <sup>3</sup> *Department of Ecology and Evolutionary Biology, University of Toronto, 25 Willcocks*  
11                  *Street, Toronto, Ontario, Canada M5S 3B2*

12  
13                  \*Author for correspondence: Joana Costa, Tel.: +351936936456, E-mail  
14                  [jfmc.biologia@gmail.com](mailto:jfmc.biologia@gmail.com)

15  
16                  Total word count: 6105

17                  Introduction: 1251 words, Materials and Methods: 1702 words, Results: 768 words,

18                  Discussion: 2280 words, Acknowledgments: 104 words

19                  Number of figures: 3 (Figures 1 and 3 to be published in colour), number of tables: 2,

20                  Supporting Information: 8 files

## 28 **Summary**

- 29 ● The evolutionary pathways leading to the heterostylous syndrome are not well  
30 understood and models concerning the origins of distyly differ in the order in which  
31 reciprocal herkogamy and self-incompatibility evolve. We investigated the evolution  
32 and breakdown of distyly in Plumbaginaceae, a family with considerable diversity of  
33 floral traits and reproductive systems.
- 34 ● Using Bayesian Markov chain Monte Carlo analyses and stochastic character  
35 mapping, we examined the evolutionary assembly and breakdown of the  
36 heterostylous syndrome based on a well-resolved phylogeny of 121 species of  
37 Plumbaginaceae and six outgroup taxa using five nuclear and plastid gene regions.  
38 We used the distribution of reproductive traits and reconstructed ancestral characters  
39 across phylogenies to evaluate competing models for the evolution of distyly.
- 40 ● The most likely common ancestor of Plumbaginaceae was self-incompatible and  
41 monomorphic for sex-organ arrangement and pollen-stigma characters. Character  
42 state reconstructions indicated that reciprocal herkogamy evolved at least three times  
43 and that shifts to selfing and apomixis occurred on multiple occasions.
- 44 ● Our results provide comparative support for the early ideas of H.G. Baker on  
45 evolutionary pathways in Plumbaginaceae, and the more recent selfing avoidance  
46 model by D. & B. Charlesworth in which distyly evolves from self-incompatible  
47 ancestors.

48

49 **Key words:** ancestral state reconstructions, heteromorphic self-incompatibility, floral  
50 evolution, heterostyly, pollen-stigma dimorphism, Plumbaginaceae, reciprocal  
51 herkogamy.

52

53

54

55

56

57

## 58 **Introduction**

59 Heterostyly is a remarkable example of a convergent floral polymorphism that has  
60 evolved numerous times in at least 28 angiosperm families (Ganders, 1979; Lloyd &  
61 Webb, 1992a; Barrett & Shore, 2008). Heterostylous populations have a reciprocal  
62 arrangement of stigma and anther heights, a condition referred to as reciprocal  
63 herkogamy (Webb & Lloyd, 1986), and are usually comprised of two (distyly) or three  
64 (tristyly) style morphs. The polymorphism in stigma and anther height is usually  
65 accompanied by a heteromorphic incompatibility system under sporophytic control, and  
66 by ancillary polymorphisms of stigmatic papillae and pollen. The evolution and  
67 breakdown of heterostyly has received sustained interest since Darwin's (1877) classic  
68 book on floral polymorphisms (reviewed in Barrett, 1992; Weller, 2009; Cohen, 2010).  
69 However, it was not until the latter part of the last century that evolutionary biologists  
70 began to investigate the evolutionary buildup of this floral syndrome using theoretical  
71 models (Charlesworth & Charlesworth, 1979; Lloyd & Webb, 1992b). The models  
72 differ in the sequence in which the morphological characters and physiological  
73 incompatibility system are established, and also in the emphasis placed on various  
74 selective forces.

75 Charlesworth & Charlesworth's (1979) 'selfing avoidance' model postulates that  
76 the ancestral condition for distyly involves a monomorphic population with plants  
77 possessing long styles and long-level anthers. Self-incompatibility with two mating  
78 types (often referred to as diallelic or heteromorphic incompatibility) establishes first to  
79 avoid selfing and inbreeding depression, with reciprocal herkogamy subsequently  
80 evolving to improve cross-pollination between the two cross-compatible mating types.  
81 The second step in this model is often described as "Darwin's cross-promotion  
82 hypothesis" (Darwin, 1877). By contrast, the 'pollen transfer' model of Lloyd & Webb  
83 (1992b) assumes that the promotion of cross-pollination is the principal selective force  
84 for the evolution of distyly, as Darwin (1877) proposed. In their model, the ancestral  
85 condition is approach herkogamy (stigmas positioned above anthers in flowers) and  
86 reciprocal herkogamy establishes to promote more proficient cross-pollen transfer  
87 among plants in the population. Subsequently, heteromorphic incompatibility may or  
88 may not evolve, depending on the severity of inbreeding depression. In both models,  
89 stigma-height dimorphism is an intermediate and transitory stage.

90 Comparative data from phylogenetic reconstructions and character mapping of a  
 91 few heterostylous lineages have provided some support for various aspects of Lloyd and  
 92 Webb's model (e.g., *Narcissus*, Graham & Barrett, 2004; *Lithodora* and related  
 93 *Glandora*, Ferrero *et al.*, 2009a, 2012; *Exochaenium*, Kissling & Barrett, 2013). In  
 94 addition, experimental work on pollination (Stone & Thomson, 1994; Cesaro &  
 95 Thompson, 2004; Keller *et al.*, 2014) and mating patterns using genetic markers (Kohn  
 96 & Barrett, 1992; Zhou *et al.*, 2015) have also provided support for Darwin's cross-  
 97 promotion hypothesis but do not allow rejection of either model because both involve  
 98 this selective force at different stages in the buildup of the polymorphism. Because  
 99 heterostyly has clearly evolved on numerous occasions, it is possible that both models  
 100 for the evolution of distyly will be supported once more data become available from  
 101 additional families.

102 Plumbaginaceae is of worldwide distribution and is comprised of 27 genera and  
 103 c. 650 species (Kubitzki, 1993). The family exhibits great diversity in morphological  
 104 characters associated with the heterostylous syndrome, thus providing an opportunity to  
 105 investigate the evolutionary buildup and breakdown of floral polymorphisms. This was  
 106 recognized by the renowned plant evolutionist Herbert G. Baker (1948a,b, 1966), who  
 107 pointed out that species from subfamily Staticoideae displayed significant  
 108 morphological differences in pollen and stigma characters when compared with those in  
 109 subfamily Plumbaginoideae. Taxa in Plumbaginoideae are largely distylous, self-  
 110 incompatible and display relatively uniform patterns of pollen exine ornamentation and  
 111 stigmatic papillae shape between style morphs (Baker, 1948a, 1966; Dulberger, 1975a;  
 112 Erdtman, 1986; Ghobary, 1986; Ferrero *et al.*, 2009b). By contrast, species in subfamily  
 113 Staticoideae exhibit considerable morphological variation and particularly striking  
 114 pollen-stigma dimorphisms (i.e., ancillary pollen and stigma characters).

115 Generally referred to as *Armeria*-type pollen, as opposed to *Plumbago*-type  
 116 pollen from the Plumbaginoideae (Erdtman, 1986), dimorphic pollen in Staticoideae  
 117 commonly has a coarse or finely reticulate sexine, termed pollen type A or B,  
 118 respectively. Plants producing pollen type A have cob-like stigmatic papillae, whereas  
 119 plants with pollen type B have papillate stigmas (Baker, 1948a,b, 1966; Dulberger,  
 120 1975a,b, 1992; and see Figs. 1, 2 in Costa *et al.*, 2017). In common with other  
 121 heterostylous plants (reviewed in Dulberger, 1992), pollen-stigma dimorphism in the  
 122 Plumbaginaceae is very closely associated with heteromorphic self-incompatibility. In  
 123 Staticoideae, where the dimorphisms are especially well developed, they presumably

124 function to promote disassortative pollination because of the absence of reciprocal  
125 herkogamy (see Dulberger, 1975a; Mattsson, 1983; Dulberger, 1992), and indeed there  
126 is experimental evidence supporting this hypothesis (Mattsson, 1983; Costa *et al.*,  
127 2017).

128         The first detailed hypothesis for the buildup and breakdown of the heterostylous  
129 syndrome in Plumbaginaceae was proposed by Baker (1948a, 1966), based on patterns  
130 of geographical distribution of species and studies of variation in morphological  
131 characters, particularly of pollen and stigmas. However, Baker's hypothesis was not  
132 developed from explicit phylogenetic methods; he simply ordered species and  
133 characters in a particular sequence, based on his own ideas on the most plausible  
134 evolutionary pathways (e.g., Baker, 1966). He postulated that the heterostylous  
135 syndrome evolved from a common ancestor with heteromorphic incompatibility and  
136 monomorphic pollen and stigmas, but by different pathways in the two subfamilies  
137 (Baker, 1948a, 1966).

138         In Plumbaginoideae, Baker proposed that reciprocal herkogamy was the only  
139 morphological addition to the ancestral incompatibility system, whereas in Staticoideae,  
140 the floral polymorphisms evolved in three stages. Specifically, he proposed that pollen  
141 dimorphism established first in species with monomorphic capitate stigmas, as in  
142 *Goniolimon*, *Acantholimon* and some *Limonium*. Subsequently, filiform dimorphic  
143 stigmas and dimorphic pollen evolved in species of *Armeria*, *Limoniastrum* and  
144 *Limonium*. Finally, reciprocal herkogamy evolved in the pollen-stigma dimorphic  
145 *Limonium vulgare*. Baker assessed the compatibility status of a large number of species  
146 of *Limonium* by performing controlled hand-pollinations (Baker, 1953a,b) and inferred  
147 that self-incompatibility was widespread in the family, apart from a few self-compatible  
148 or apomictic species of *Limonium* with “secondary monomorphism” of pollen and  
149 stigmas. He considered this secondary monomorphism as derived from pollen-stigma  
150 dimorphism as a result of either the evolution of homostyly [small-flowered selfing  
151 forms derived from heterostylous ancestors (see Darwin, 1877; Ganders, 1979)], or  
152 through the replacement of sexual reproduction by apomixis (Baker, 1966).

153         Significantly, Baker's hypothesis (Baker, 1948a, 1966) shares features in common with  
154 the model of D. & B. Charlesworth (1979) for the evolution of distyly as both involve  
155 the evolution of self-incompatibility prior to the establishment of reciprocal herkogamy.

156         Here, we investigate Baker's ideas on the evolutionary buildup and breakdown  
157 of the heterostylous syndrome in Plumbaginaceae using models of character evolution

158 and stochastic character mapping. We used molecular data to construct a phylogenetic  
159 tree for the family and an extensive survey of herbarium specimens and the literature to  
160 investigate the evolution of heteromorphic morphological traits. Our study addressed  
161 the following specific questions: (1) Was the most likely common ancestor of  
162 Plumbaginaceae self-incompatible with monomorphic pollen and stigmas, as Baker  
163 (1948a, 1966) originally proposed? (2) How many times has reciprocal herkogamy  
164 evolved in the family and was this prior to or after the establishment of self-  
165 incompatibility and pollen and stigma dimorphisms? (3) Is there phylogenetic evidence  
166 of breakdown of the distylous syndrome to alternative reproductive systems involving  
167 homostyly and apomixis? (4) Is reciprocal herkogamy more likely to be selected in  
168 lineages with pollen-stigma dimorphism? (5) Does apomixis evolve following the loss  
169 of pollen-stigma dimorphism in subfamily Staticoideae? Our results provide support for  
170 several of Baker's proposals and also for the order in which self-incompatibility and  
171 reciprocal herkogamy are established in the Charlesworth and Charlesworth's (1979)  
172 model of the evolution of distyly.

173

## 174 **Materials and Methods**

### 175 **Characterization of floral polymorphisms and associated characters**

176 To characterize the distribution of reproductive traits among taxa of Plumbaginaceae,  
177 we combined extensive literature searches and sampling of herbarium specimens  
178 (number of specimens per species: mean  $\pm$  SD  $3.10 \pm 1.23$ , median 3, range 1-6) for a  
179 total of 121 species. Details are provided in Supporting Information Methods 1 and the  
180 herbarium specimens sampled are listed in Supporting Information Table S1.  
181 Information from the literature was available for 61 species in the dataset and for 33 this  
182 was combined with herbarium sampling. The characterization of floral polymorphisms  
183 for 59 species was based on herbarium sampling only. The resulting character matrix  
184 contained six traits (see Supporting Information Table S2): sex-organ arrangement  
185 (reciprocal herkogamy vs style-stamen height monomorphism), incompatibility status  
186 (self-incompatibility vs self-compatibility), exine sculpturing (*Armeria*-type pollen vs  
187 monomorphic pollen), stigmatic papillae condition (stigma dimorphism vs stigma  
188 monomorphism), stigma morphology (filiform vs capitate vs others) and reproductive  
189 mode (sexual vs apomixis).

190 We followed several criteria in constructing the character matrix: (1) Because  
191 most species included in the analyses were not tested experimentally for their  
192 incompatibility status, we used two approaches to infer whether they were self-  
193 incompatible or self-compatible. For species in Plumbaginoideae, data on pollen type is  
194 uninformative with regards to incompatibility status and thus we used information in the  
195 literature, whereas for Staticoideae information from the literature (e.g., Baker, 1966)  
196 was combined with microscopic observations of pollen exine sculpturing and stigmatic  
197 papillae shape from herbarium specimens. Species were scored as self-incompatible if  
198 they possessed A/cob and B/papillate pollen-stigma combinations, and as self-  
199 compatible if they had A/papillate or B/cob pollen-stigma combinations. This approach  
200 follows criteria established by Baker (1948a, 1966) based on controlled pollinations and  
201 also Erben (1979). (2) As *Plumbago*-type pollen shows only very slight differences  
202 between style morphs in exine sculpturing (e.g., Ghobary, 1986; Ferrero *et al.*, 2009b),  
203 this condition was considered monomorphic in all analyses. (3) Species of *Limonium*  
204 were scored as sexual or apomictic based on observations of pollen exine sculpturing  
205 and stigmatic papillae shape combined with information on chromosome counts from  
206 the literature. Dimorphic species with A/cob and B/papillate pollen-stigma  
207 combinations and even chromosome counts of  $2n = 12, 14, 16, 18, 32, 34, 36, 54$  were  
208 considered sexual, whereas species monomorphic for pollen and stigmas (A/cob or  
209 B/papillate) with odd chromosome counts were scored as apomictic. This approach also  
210 follows Erben (1979) and data available from Baker (1966). We confirmed and assigned  
211 all species names following *The Plant List* (2013) and *Species 2000 & ITIS Catalogue*  
212 *of Life* (Roskov *et al.*, 2016) available online.

213

## 214 Phylogenetic analyses

215 We used a supermatrix approach (de Queiroz & Gatesy, 2007; Roquet *et al.*, 2013) to  
216 construct a phylogenetic hypothesis for evolutionary relationships of species in the  
217 ingroup. Sequences of the five most common markers used in previous phylogenetic  
218 studies of Plumbaginaceae were downloaded from GenBank using Geneious v9.0.5  
219 (Kearse *et al.*, 2012). We used one marker of the nuclear genome (*ITS*) and four of the  
220 plastid genome (*rbcL*, *matK*, intergenic spacers *trnL-trnF* and *trnT-trnY*) (Supporting  
221 Information Table S3). Our outgroup included six species from three families  
222 considered to be the closest relatives of Plumbaginaceae: *Polygonum amphibium*, *P.*



223 *capitatum*, *Rumex obtusifolius* and *Triplaris americana* (Polygonaceae), *Tamarix*  
 224 *aphylla* (Tamaricaceae) and *Frankenia pulverulenta* (Frankeniaceae) (Soltis *et al.*,  
 225 2000).

226 We aligned sequences using MAFFT v7 available online  
 227 (<https://mafft.cbrc.jp/alignment/server/>; Katoh & Standley, 2013). All alignments were  
 228 improved by removing poorly aligned or ambiguous regions by setting “automated 1”  
 229 function on trimAl v1.3 software (Capella-Gutiérrez *et al.*, 2009) available online at the  
 230 Phylemon 2.0 server (<http://phylemon.bioinfo.cipf.es/index.html>; Sánchez *et al.*, 2011).  
 231 To construct the supermatrix, trimmed alignments were concatenated by using  
 232 FASconCAT v1.0 (Kück & Meusemann, 2010) resulting in a supermatrix of 4480  
 233 characters and 94 taxa. We analyzed the supermatrix using a maximum likelihood (ML)  
 234 approach following the GTR model as implemented in RAxML v8.2.8 (Stamatakis,  
 235 2014) available at the CIPRES server (<http://phylo.org>, Miller *et al.*, 2010) by running  
 236 1000 bootstrap replicates. We kept the best ML phylogenetic tree and nodes with  
 237 bootstrap values lower than 50% were collapsed using TreeGraph 2 (Stöver & Müller,  
 238 2010).

239 To obtain an ultrametric phylogenetic tree, we used molecular dating by  
 240 likelihood methods as implemented in the *chronos* function available from the R  
 241 package “ape” (Paradis *et al.*, 2004; Paradis, 2013) on the ML tree previously obtained.  
 242 We obtained minimum and maximum ages for Plumbaginaceae from Bell *et al.* (2010)  
 243 and for calibration purposes used minimum = 27 mya and maximum = 57 mya. The  
 244 selection of the model of substitution rate was based on the information criterion PHIIC  
 245 and the model with the lowest PHIIC was selected (Paradis, 2013). PHIIC values for the  
 246 tested models are available in Supporting Information Table S4. There were no  
 247 differences when changing the smoothing parameter for the model “strict” and therefore  
 248 the simplest one was chosen (model “strict”, lambda value “0”).

249 For 35 species included in the character matrix for which there were no  
 250 sequences available in GenBank, we searched in the literature for information regarding  
 251 monophyly of the genus. If that was the case, species were grafted onto their respective  
 252 genera in the ultrametric tree by using *add.species.to.genus* function from R package  
 253 “phytools” (Revell, 2012). When no information regarding monophyly was available,  
 254 we searched the literature for the putatively closest relative and added the species to a  
 255 specific node by setting *bind.tip* function from R package “phytools” (Revell, 2012).  
 256 For *Limonium minutiflorum*, the closest relatives [i.e., species from the same section



257 according to Boissier (1848)] were in two different positions in the tree (Supporting  
258 Information Fig. S1). To reduce inaccuracies in character reconstructions associated  
259 with the grafting of this species, two trees each with *L. minutiflorum* in a different  
260 position were created. The grafting of species introduced some polytomies in the final  
261 trees that were randomly resolved by creating two sets of 100 trees using Mesquite  
262 v3.04 (Maddison & Maddison, 2015). The ingroup included 121 species, seven of  
263 Plumbaginoideae and 114 species of Staticoideae, from 14 genera representing ~19% of  
264 the total species in Plumbaginaceae.

265

## 266 Inferences on ancestral states and character mapping

267 To assess the ancestral states of the six characters investigated, we conducted Bayesian  
268 Markov chain Monte Carlo (MCMC) analyses in two sets of 100 trees (see *Phylogenetic*  
269 *analyses*), thus accounting for phylogenetic uncertainty. We used the MultiState  
270 continuous-time Markov model of character evolution for discrete data as implemented  
271 in BayesTraits v2 (Pagel *et al.*, 2004). To reduce the uncertainty and randomness  
272 associated with setting the prior distribution, we used hyperpriors as recommended by  
273 Pagel *et al.* (2004). The hyperprior is a uniform distribution that is used to seed values  
274 of the gamma or exponential priors (Pagel *et al.*, 2004).

275 To test for the most likely ancestral character states, we implemented MCMC  
276 analyses by running Markov chains for one million iterations that were sampled each  
277 1000 steps, with a burn-in of 10 000, and by setting the stepping stone sampling to use  
278 100 stones and run each for 10 000 iterations. The node containing all species of  
279 Plumbaginaceae was fixed to each possible state of a given character under two models  
280 of transition rates between character states: (1) equal rates (ER), and (2) all rates  
281 different (ARD). For incompatibility status and reproductive mode, two additional  
282 analyses were run by restricting back transitions (i.e., SC to SI and apomixis to sexual)  
283 to zero. In total, we tested four models for each trait (five for incompatibility status and  
284 reproductive mode, six for stigma morphology), and the best fitting model was selected  
285 based on the Bayesian Information Criterion (BIC).

286 We mapped transitions between states for the six characters by means of  
287 stochastic character mapping (SCM; Huelsenbeck *et al.*, 2003) in the two sets of 100  
288 trees using *make.simmap* function from R package “phytools” (Revell, 2012). This  
289 function simulates stochastic character histories on a phylogenetic tree or sets of trees.

290 We used the most likely value of the transition matrix to fit the prior distribution, except  
 291 for incompatibility status and reproductive mode. The loss of complex characters such  
 292 as these is generally irreversible (Dollo, 1893; Bull & Charnov, 1985; Goldberg & Igić,  
 293 2008), and thus low transition rates were defined in the Q matrix (SI to SC, sexual to  
 294 apomictic:  $1E^{-3}$ ; SC to SI, apomictic to sexual:  $1E^{-5}$ ). The best fitting model of character  
 295 evolution (ER vs ARD) as revealed by MCMC analyses was used for each trait. We  
 296 performed Markov chains for 100 simulations for each tree, that is 10 000 simulations  
 297 for each set of trees for each character. Because the *make.simmap* function allows  
 298 estimation of the posterior probability (PP) of all possible states for a character that is  
 299 considered uncertain, we used a binary matrix, i.e., a matrix of probabilities ranging  
 300 from 0 to 1 for each trait. Species with missing data for a given character were assumed  
 301 to have equal prior probability of taking one or the other character state.

302 Character mapping allows the estimation of the PP of states of the most recent  
 303 common ancestor of each node, and the calculation of the average number of changes  
 304 between states, as well as the time spent in each state. We obtained the relative rate of  
 305 transition between states by dividing the frequency of each transition (i.e., the ratio  
 306 between one specific change and the total number of changes) by the frequency of time  
 307 spent in each state (Torices & Anderberg, 2009).

308

### 309 Correlated evolution between characters

310 To test for: (1) correlated evolution between reciprocal herkogamy and pollen-stigma  
 311 dimorphism, and (2) whether apomixis originates following the loss of pollen-stigma  
 312 dimorphism, we conducted MCMC analyses using *Discrete independent* (traits are  
 313 assumed to evolve independently) and *Discrete dependent* (the rate of change of one  
 314 trait depends on the state of the other) options as implemented in BayesTraits v2 (Pagel  
 315 *et al.*, 2004). We scored species as pollen-stigma dimorphic based on the possession of  
 316 pollen and stigmatic papillae dimorphism simultaneously. The stepping stone sampling  
 317 was set to use 100 stones and run each for 10 000 iterations. Under these models,  
 318 transitions between states are only allowed for one character at each time. We compared  
 319 the likelihoods of the two models by means of Bayes Factor (BF) and values  $\text{Log BF} >$   
 320 2 indicated evidence for correlated evolution.

321

## 322 Results

### 323 Phylogenetic distribution of character states

324 Reciprocal herkogamy occurs in the two subfamilies of Plumbaginaceae, but its overall  
 325 frequency differs between them (Fig. 1, Supporting Information Table S2). All species  
 326 of Plumbaginoideae in the dataset ( $n = 7$ ) possess reciprocal herkogamy and are  
 327 therefore distylous. In the Staticoideae, distyly occurs in *Goniolimon*, some  
 328 *Acantholimon* species and *Limonium vulgare* for a total of 12 species of the 114  
 329 investigated in the subfamily. All species of Plumbaginoideae in the dataset were self-  
 330 incompatible ( $n = 7$ ), whereas self-incompatibility, as inferred based on pollen-stigma  
 331 character state combinations, occurred in 101 species of Staticoideae. Self-compatibility  
 332 was inferred in four species, all *Limonium*, in subfamily Staticoideae, and nine species  
 333 had missing data for this trait (Supporting Information Table S2).

334 Regarding ancillary polymorphisms of pollen and stigma (Fig. 1, Supporting  
 335 Information Table S2), *Armeria*-type pollen was only found in Staticoideae and  
 336 monomorphic exine of the *Plumbago*-type appeared to be restricted to Plumbaginoideae  
 337 ( $n = 7$ ). However, eight species of *Limonium* produced only one pollen type, mainly  
 338 type A (*L. echioides* produced pollen type B) and these species were inferred to possess  
 339 monomorphic pollen. Similarly, stigma dimorphism only occurred in species of  
 340 subfamily Staticoideae and 11 species of *Limonium* had monomorphic stigmas, which  
 341 were either cob or papillate. Capitate stigmas were characteristic of all Plumbaginoideae  
 342 species included in this study, and also species of *Acantholimon* ( $n = 38$ ),  
 343 *Cephalorrhizum* ( $n = 1$ ), *Dictyolimon* ( $n = 1$ ) and *Goniolimon* ( $n = 3$ ), all in the  
 344 Staticoideae (Fig. 1, Supporting Information Table S2).

345 Almost all species investigated were inferred to be sexual based on data from the  
 346 literature and the combination of pollen-stigma dimorphism with chromosome counts.  
 347 A few *Limonium* species ( $n = 8$ ) were inferred to be apomictic as they exhibited  
 348 monomorphic pollen and stigmas and odd-numbered chromosomes (Fig. 1, Supporting  
 349 Information Table S2). Species of *Aegialitis* shared few of the character states with  
 350 other species of Staticoideae. *Aegialitis annulata* and *A. rotundifolia* lacked reciprocal  
 351 herkogamy, possessed monomorphic pollen and capitate stigmas and were inferred to  
 352 be sexual (Fig. 1, Supporting Information Table S2).

353

### 354 Character evolution

355 The results obtained from MCMC analyses and SCM, using the two sets of 100 trees  
 356 differing in the position of *Limonium minutiflorum*, were very similar with respect to  
 357 the most likely ancestral condition of Plumbaginaceae and the number of transitions  
 358 between character states. Therefore, only results for one set of trees are described  
 359 below. Results for the second set are provided in Supporting Information Tables S5-S7.

360 Figure 2 illustrates the most likely pathways of reproductive character evolution  
 361 in Plumbaginaceae revealed by our analyses. The most recent common ancestor of  
 362 Plumbaginaceae was most likely monomorphic for sex-organ arrangement, self-  
 363 incompatible, pollen and stigma monomorphic with capitate stigmas and was sexual  
 364 (Table 1). Our analysis indicates that reciprocal herkogamy evolved at least three times  
 365 in the family (Table 2), and was the most likely condition of the most recent common  
 366 ancestor of species in subfamily Plumbaginoideae and the clade containing genera  
 367 *Acantholimon* and *Goniolimon* (Fig. 1). In addition, the most likely change in sex-organ  
 368 arrangement was the breakdown of reciprocal herkogamy to homostyly (relative rate of  
 369 transition: 1.80). The loss of incompatibility was estimated to have occurred at least five  
 370 times ( $4.99 \pm 0.004$ ) in the family.

371 The average number of transitions ( $\pm$  SE) from the ancestral monomorphic state  
 372 to dimorphic pollen and stigmas was  $2.39 \pm 0.01$  and  $7.55 \pm 0.03$ , respectively. The  
 373 opposite pattern, involving reversion from dimorphism to monomorphism for pollen  
 374 and stigmas was estimated to be  $9.28 \pm 0.02$  and  $11.08 \pm 0.03$ , respectively. The loss of  
 375 pollen dimorphism was estimated to be three times more likely to occur than the  
 376 opposite transition, whereas the loss of stigmatic papillae dimorphisms was only  
 377 slightly larger than their gain in the Staticoideae (Table 2). The transition from capitate  
 378 to filiform stigmas most likely occurred only once ( $1.37 \pm 0.01$ ), whereas the reversion  
 379 was found to be even less likely (Table 2). The change in reproductive mode from  
 380 sexual to asexual was restricted to *Limonium* species (Fig. 1) and we estimate that this  
 381 transition most likely occurred five times ( $5.01 \pm 0.02$ ).

382

### 383 Correlated evolution between characters

384 We found evidence that reciprocal herkogamy and pollen-stigma dimorphism evolved  
 385 independently of one another in Plumbaginaceae (BF = -21.111). In contrast, we  
 386 detected a strong correlation between the joint evolution of reproductive mode and  
 387 pollen-stigma dimorphism (BF = 12.771; Fig. 3). In subfamily Staticoideae, the shift

388 from sexual to apomictic reproduction ( $q_{24} = 0.0179$ ) appears to precede the loss of  
389 pollen-stigma dimorphism ( $q_{21} = 0.0112$ ) and most likely involved an intermediate stage  
390 of apomixis with pollen-stigma dimorphism.

391

## 392 **Discussion**

393 Our comparative analysis of the evolutionary history of the origin and breakdown of the  
394 heterostylous syndrome in Plumbaginaceae revealed several main findings: (1) The  
395 most recent common ancestor of the family was likely self-incompatible with a uniform  
396 arrangement of sex organs and monomorphic pollen and stigmas. (2) Reciprocal  
397 herkogamy evolved on at least three different occasions from an ancestor with  
398 monomorphic style and stamen lengths. (3) Pollen-stigma dimorphism is likely to have  
399 evolved before the establishment of reciprocal herkogamy in Statioideae. (4)  
400 Transitions from pollen-stigma dimorphism to monomorphism, associated with the  
401 evolution of self-compatibility and apomixis, occurred on multiple occasions in  
402 *Limonium*. Our study is the first comprehensive re-evaluation of Baker's hypotheses for  
403 the evolutionary buildup and breakdown of the heterostylous syndrome in  
404 Plumbaginaceae. It provides some support for Baker's proposals and more broadly to  
405 aspects of the selfing avoidance model for the evolution of distyly (Charlesworth &  
406 Charlesworth, 1979; Fig. 2). Below, we discuss our findings and provide several  
407 refinements to the evolutionary pathways originally proposed by Baker (1948a, 1966).

408

### 409 Evolution of reciprocal herkogamy

410 The results of stochastic character mapping indicated that reciprocal herkogamy evolved  
411 on at least three occasions from a common ancestor with a monomorphic arrangement  
412 of sex organs; once in Plumbaginoideae and twice in Statioideae. Earlier phylogenetic  
413 reconstructions of heterostylous groups have provided evidence for multiple origins of  
414 reciprocal herkogamy in various taxa including including Boraginaceae (Ferrero *et al.*,  
415 2009a; Cohen, 2013), *Linum* (Armbruster *et al.*, 2006; McDill *et al.*, 2009), *Narcissus*  
416 (Pérez *et al.*, 2003; Graham & Barrett, 2004) and *Nymphoides* (Tippery & Les, 2011).  
417 Concerning Plumbaginoideae, we included species from three of the four currently  
418 accepted genera – *Plumbago*, *Dyerophytum* and *Ceratostigma* – and all investigated  
419 species in these genera were distylous. Evidence from SCM suggested that the ancestral  
420 condition in this subfamily is reciprocal herkogamy. The fourth genus in

421 Plumbaginoideae is *Plumbagella*, which is Asian in distribution and is comprised of a  
 422 single species, *P. micrantha* (Kubitzki, 1993). This species is annual, the corolla is very  
 423 reduced in size (ca. 4-5 mm; eFloras, 2008) and its monomorphic arrangement of sex  
 424 organs is most probably a derived condition involving the evolutionary breakdown of  
 425 distyly to homostyly (see Baker, 1948a, 1966), rather than an ancestral monomorphic  
 426 condition. In common with other homostylous taxa in heterostylous groups (Darwin,  
 427 1877; reviewed in Ganders, 1979; Barrett, 1989), *P. micrantha* is probably autogamous  
 428 and the facility for autonomous self-pollination may have enabled establishment  
 429 following long-distance dispersal to Asia.

430 The most likely common ancestor of subfamily Plumbaginoideae had a  
 431 reciprocal arrangement of sex-organs, whereas the common ancestor of all Staticoideae  
 432 most likely possessed a monomorphic arrangement of sex organs, although this does not  
 433 imply that they were necessarily selfing, as seems likely for derived homostylous taxa  
 434 in the family. Reciprocal herkogamy appears to have evolved at least twice from stylar  
 435 monomorphism in Staticoideae. *Limonium vulgare* was originally described as distylous  
 436 and this was thought to be the only case of reciprocal herkogamy in Staticoideae (Baker,  
 437 1948a, 1966). Our results support the hypothesis that reciprocal herkogamy is derived  
 438 from style-stamen monomorphism (and see Baker, 1948a, 1966). However, later studies  
 439 of *Acantholimon* (Bokhari, 1972), *Goniolimon tataricum* (Schill *et al.*, 1985) and *G.*  
 440 *italicum* (Morretti *et al.*, 2015), as well as our own sampling of herbarium specimens,  
 441 demonstrate that the occurrence of distyly is more widespread in Staticoideae than  
 442 originally thought, occurring in at least 12 species. *Acantholimon* and *Goniolimon* form  
 443 a clade (Fig. 1; and also see Lledó *et al.*, 1998, 2001, 2005) for which reciprocal  
 444 herkogamy is the most probable basal condition, as revealed by SCM. The lack of  
 445 reciprocal herkogamy in some species of *Acantholimon* is therefore likely to be a  
 446 derived condition, and these taxa are probably homostylous resulting from the  
 447 breakdown of distyly, as inferred for *Plumbagella*.

448

#### 449 Pollen-stigma dimorphism and incompatibility

450 The most likely common ancestor of Plumbaginaceae had monomorphic pollen, capitate  
 451 stigmas and was self-incompatible. To infer the compatibility status of species in  
 452 Plumbaginaceae, we combined experimental evidence available from controlled hand  
 453 pollinations (e.g., Baker, 1953b; Dulberger, 1975a; Ferrero *et al.*, 2009b; reviewed in



454 Dulberger, 1992), and assumptions based on pollen exine ornamentation and stigmatic  
455 papillae shape in Staticoideae. Species of Plumbaginoideae resemble the basal condition  
456 in the family in having no obvious signs of pollen-stigma dimorphism and capitate  
457 stigmas (Fig. 1, Supporting Information Table S2; Baker, 1948a,b, 1966). Although  
458 there are very slight differences in exine sculpturing, as revealed by scanning electron  
459 microscopy of pollen from the long- and short-styled morphs of *Dyerophyton africanum*  
460 and *D. indicum* (Ghobary, 1986), these have not been considered sufficiently well  
461 developed to classify *Plumbago*-type pollen as dimorphic (Erdtman, 1986).  
462 Experimental tests for the presence of self-incompatibility have been conducted in some  
463 species of Plumbaginoideae [e.g., *Plumbago capensis* and *P. europaea* (Dulberger,  
464 1975a); *P. auriculata* (Ferrero *et al.*, 2009b); *Ceratostigma willmottianum* (Dulberger,  
465 1975a)], and with the exception of *Plumbagella micrantha* (Dahlgren, 1918), they were  
466 all found to be self-incompatible setting no or few seeds upon self-pollination. As  
467 mentioned above, self-compatibility in *P. micrantha* is considered a derived condition  
468 allowing reproduction by seed in this annual, likely homostylous, species (see Baker,  
469 1948a).

470 In contrast, pollen-stigma dimorphism is highly developed in Staticoideae,  
471 except for *Aegialitis*. The only two species in this genus, *A. annulata* and *A.*  
472 *rotundifolia*, are distinct from the remaining Staticoideae in having *Plumbago*-type  
473 pollen (Baker, 1966; Ghobary, 1984) and monomorphic capitate stigmas (Baker, 1966;  
474 Lledó *et al.*, 2001). Indeed, this genus is probably one of the earliest branching lineages  
475 in Staticoideae (Baker, 1948a,b, 1966). More recently, the phylogenetic position of  
476 *Aegialitis* has been investigated by parsimony analyses and the conclusion reached that  
477 the genus constitutes a distinct lineage within Staticoideae (Lledó *et al.*, 2001), which  
478 has diverged from the most likely common ancestor before pollen-stigma dimorphism  
479 evolved in the subfamily (and see Baker, 1948a,b, 1966).

480 Stochastic character mapping revealed a single origin of pollen-stigma  
481 dimorphism in Plumbaginaceae that was “superimposed” on a previously established  
482 self-incompatibility system. This may have occurred soon after the divergence of the  
483 clade containing *Aegialitis* in Staticoideae. Investigations in *Limonium* (Baker, 1953a;  
484 Dulberger, 1975a) and *Armeria maritima* (Iversen, 1940; Dulberger, 1975a; Mattsson,  
485 1983; Costa *et al.*, 2017) have indicated that species with pollen-stigma dimorphism  
486 possess heteromorphic incompatibility, a widespread association in most heterostylous  
487 families (Dulberger, 1992). In addition to marked differences in exine sculpturing,



488 pollen types A and B also differ in their lipid composition (Mattsson, 1983), which  
489 probably plays a key role in the initial interaction between pollen and stigmas, during  
490 the adhesion phase (Mattsson, 1983; Heslop-Harrison & Heslop-Harrison, 1985).  
491 Heteromorphic incompatibility and pollen and stigma polymorphisms are tightly  
492 associated in this subfamily and both physiological and morphological characters  
493 appear to participate in the rejection of self-pollen (Dulberger, 1975a,b; Costa *et al.*,  
494 2017), a situation that does not occur in species with homomorphic incompatibility  
495 (reviewed in Franklin-Tong, 2008), where *S*-gene protein specificities govern self-  
496 rejection.

497

#### 498 The breakdown of heterostyly to uniparental reproduction

499 The shift from obligate outcrossing to predominant selfing is generally considered the  
500 most common mating-system transition in angiosperms (Stebbins, 1974), with selfing  
501 commonly selected when it confers reproductive assurance (Lloyd, 1992), especially  
502 after long-distance dispersal and during repeated colonization episodes (Baker, 1955,  
503 1959, 1967; Pannell *et al.*, 2015). Our analysis of the distribution of characters on  
504 phylogenetic trees suggests that the transition from pollen-stigma dimorphism and self-  
505 incompatibility to monomorphism and self-compatibility most likely occurred at least  
506 four times independently in *Limonium* (Fig. 1). In addition, pollen-stigma  
507 monomorphism and self-compatibility are also reported for *Armeria maritima* subsp.  
508 *sibirica* (Baker, 1948c, 1966) and some *Limonium* species from North America and  
509 Australia (Baker, 1953a,b), but these were not included in our character mapping.  
510 Monomorphic self-compatible species most frequently produce type A pollen and have  
511 papillate stigmas, whereas the combination B/cob has only been found in the annual  
512 Mediterranean species *L. echioides*. It is probable that in *Limonium* self-compatibility  
513 has arisen in association with dispersal events from Europe (Baker, 1953a,b, 1959),  
514 especially from the western Mediterranean, which is the main center of diversification  
515 of the genus (Erben, 1993; Kubitzki, 1993), to America and Australia, where species  
516 with pollen-stigma monomorphism and self-compatibility are most frequent (Baker,  
517 1953a,b). However, these hypotheses concerning migration need to be substantiated by  
518 phylogeographical analyses, which should provide valuable insights on the  
519 biogeography of reproductive transitions in this group.

520 In addition to sexual mating strategies involving outcrossing or selfing, asexual  
521 reproduction via apomixis has been described in several *Limonium* species (e.g.,  
522 D'Amato, 1940, 1949; Baker, 1953a, 1966; Erben, 1979; Ingrouille & Stace, 1985;  
523 Cowan *et al.*, 1998; Róis *et al.*, 2016). Apomictic species are most commonly polyploid  
524 (Asker & Jerling, 1992; Carman, 1997), which is also the case for apomictic *Limonium*  
525 (e.g., D'Amato, 1949; Erben, 1979; Cowan *et al.*, 1998; Caperta *et al.*, 2016; Róis *et al.*,  
526 2016). Apomixis is often associated with hybridization between species and can  
527 overcome the sexual sterility that is a feature of many interspecific hybrids (Asker &  
528 Jerling, 1992). Reproductive barriers between species of *Limonium* are often weak and  
529 hybridization is frequent (Erben, 1993). The shift from sexual to asexual reproduction  
530 via apomixis in Plumbaginaceae most likely preceded the loss of pollen-stigma  
531 dimorphism. High transition rates between apomictics with pollen-stigma dimorphisms  
532 and those with monomorphic pollen and stigmas suggest that this transition most likely  
533 involved an unstable intermediate condition of apomictics with dimorphic pollen and  
534 stigmas. Indeed, there is no report in the literature of this condition in Plumbaginaceae  
535 suggesting rapid loss of pollen-stigma dimorphism following shifts from sexual to  
536 apomictic reproduction. *Limonium* apomictics have monomorphic pollen and stigmas  
537 with combinations A/cob or B/papillate, which would normally involve a self-  
538 incompatible pollen-stigma combination. In addition, pollen production per flower is  
539 very low and pollen grains are mainly aborted (Baker, 1966; Erben, 1979; Cowan *et al.*,  
540 1998). As previously reported in other angiosperm taxa (e.g., Souza *et al.*, 2012; Aliyu  
541 *et al.*, 2013), the character mapping in our study revealed multiple independent origins  
542 of apomixis in lineages composed of mainly sexual species. The mixture of sexual and  
543 asexual reproductive strategies in *Limonium* may have contributed to the diversification  
544 of this genus in the western Mediterranean (Lledó *et al.*, 2005) and also to its taxonomic  
545 complexity.

546

#### 547 The evolutionary buildup of the heterostylous syndrome

548 Our findings are generally in accord with most of Baker's hypotheses (Baker, 1948a,  
549 1966) on the evolutionary buildup and breakdown of the heterostylous syndrome. Our  
550 analyses revealed two distinct pathways associated with the assembly of the  
551 heterostylous syndrome; a relatively simple one for Plumbaginoideae and a more  
552 complex pathway for Staticoideae (Figs. 1, 2). In Plumbaginoideae, reciprocal

553 herkogamy evolved in a self-incompatible ancestor without the development of pollen-  
554 stigma dimorphism (Figs. 1, 2). By contrast, in Statioideae the first step in the  
555 evolution of heterostyly was the acquisition of pollen-stigma dimorphism in a self-  
556 incompatible ancestor followed by the evolution of reciprocal herkogamy. In contrast to  
557 Statioideae, ancillary characters are not exceptionally well-developed in some  
558 heterostylous species (Dulberger, 1992), and this is the most likely reason why previous  
559 phylogenetic reconstructions of heterostylous lineages have not examined the  
560 evolutionary history of pollen-stigma dimorphisms. Our results suggest that pollen and  
561 stigma dimorphisms have evolved together in concert rather than sequentially. In  
562 contrast, Baker (1948a, 1966) proposed that pollen dimorphism evolved first in a self-  
563 incompatible species with monomorphic stigmas (e.g. *Goniolimon*, *Acantholimon* and  
564 some *Limonium*) and stigma polymorphism evolved subsequently. However, this order  
565 seems unlikely as later investigations of *Goniolimon* (Schill *et al.*, 1985; Morretti *et al.*,  
566 2015) and *Acantholimon* (Bokhari, 1972), and our own observations of herbarium  
567 specimens, have demonstrated that taxa in these genera all exhibit pollen-stigma  
568 dimorphism and we found no evidence of any species with only pollen dimorphism. In  
569 fact, pollen-stigma dimorphism is widespread in Statioideae, and with the exception of  
570 *Aegialitis*, monomorphic pollen and stigmas are most likely derived and associated with  
571 the transition from outcrossing to selfing or apomixis. Our results further suggest that  
572 reciprocal herkogamy most likely resulted from two independent origins after the  
573 establishment of pollen-stigma dimorphism in the clade formed by *Acantholimon* and  
574 *Goniolimon*, and separately in *Limonium*.

575         The findings of this study have general relevance for competing models of the  
576 evolution of distyly. One of the main findings of our reconstructions is that reciprocal  
577 herkogamy evolved after the establishment of heteromorphic self-incompatibility in  
578 Plumbaginaceae. This is the order of establishment proposed in the inbreeding  
579 avoidance model of Charlesworth & Charlesworth (1979) but is the opposite polarity to  
580 that predicted in the pollen transfer model of Lloyd & Webb (1992b). Our results  
581 therefore represent the first comprehensive comparative evidence on the evolution of  
582 distyly supporting the sequence proposed in the Charlesworth & Charlesworth (1979)  
583 model.

584         Despite the difference in polarities of the two models for the evolution of  
585 distyly, both involve a transient polymorphic stage of stigma-height dimorphism on the  
586 pathway from stylar monomorphism to distyly. Stigma-height dimorphism occurs in

587 species of *Narcissus* (Graham & Barrett, 2004), *Lithodora* and *Glandora* (Ferrero *et al.*,  
588 2009a, 2012), genera in which heterostylous species also occur; however, we found no  
589 evidence that any species of Plumbaginaceae possesses this form of stylar  
590 polymorphism. Stigma-height dimorphism is rather infrequent among distylous families  
591 and it is quite possible that this putative intermediate stage is ephemeral and is rapidly  
592 replaced once distyly evolves in a lineage (reviewed in Barrett *et al.*, 2000). Given the  
593 numerous independent origins of the heterostylous floral syndrome across angiosperm  
594 families, it is possible that features of both models of the evolution of heterostyly are  
595 correct for specific lineages. Future comparative work on the evolutionary history and  
596 molecular genetic architecture of characters in the heterostylous syndrome will  
597 undoubtedly shed light on which of the two pathways by which heterostyly evolves is  
598 more commonly followed and what reproductive and genetic features of lineages cause  
599 the particular pathway that is taken to occur.

600

### 601 **Acknowledgements**

602 The authors thank Fátima Sales at the Herbarium of the University of Coimbra, Lesley  
603 Scott and Suzanne Cubey at the Royal Botanical Garden of Edinburgh Herbarium, and  
604 José L. F. Alonso at Real Jardín Botánico Herbarium in Madrid for permits to sample  
605 specimens of Plumbaginaceae, and Sílvia Castro (CFE) and João Loureiro (CFE) for  
606 valuable discussions. This research was supported by POPH/FSE funds by the  
607 Portuguese Foundation for Science and Technology through a doctoral grant to Joana  
608 Costa (SFRH/BD/89910/2012), a Marie Skłodowska-Curie PostDoctoral Fellow (IF-  
609 GF) to Rubén Torices, and a Natural Sciences and Engineering Research Council  
610 Discovery Grant to Spencer C. H. Barrett.

611

### 612 **Author contributions**

613 J.C., R.T. and S.C.H.B. designed the research questions, J.C. conducted data collection,  
614 J.C. and R.T. performed data analyses, J.C. wrote the paper with input and revisions  
615 from all co-authors.

616

### 617 **References**

618 **Aliyu OM, Seifert M, Corral JM, Fuchs J, Sharbel TF. 2013.** Copy number variation  
619 in transcriptionally active regions of sexual and apomictic *Boechera* demonstrates

- 620 independently derived apomictic lineages. *Plant Cell* **25**: 3808–3823.
- 621 **Armbruster WS, Pérez-Barrales R, Arroyo J, Edwards ME, Vargas P. 2006.**  
622 Three-dimensional reciprocity of floral morphs in wild flax (*Linum suffruticosum*): a  
623 new twist on heterostyly. *New Phytologist* **171**: 581–590.
- 624 **Asker SE, Jerling L. 1992.** *Apomixis in plants*. Boca Raton, Florida: CR Press.
- 625 **Baker HG. 1948a.** Dimorphism and monomorphism in the Plumbaginaceae I. Survey  
626 of the family. *Annals of Botany* **12**: 207–219.
- 627 **Baker HG. 1948b.** Relationships in the Plumbaginaceae. *Nature* **161**: 400.
- 628 **Baker HG. 1948c.** Significance of pollen dimorphism in Late-Glacial *Armeria*. *Nature*  
629 **161**: 770–771.
- 630 **Baker HG. 1953a.** Dimorphism and monomorphism in the Plumbaginaceae II. Pollen  
631 and stigmata in the genus *Limonium*. *Annals of Botany* **17**: 433–445.
- 632 **Baker HG. 1953b.** Dimorphism and monomorphism in the Plumbaginaceae III.  
633 Correlation of geographical distribution patterns with dimorphism and  
634 monomorphism in *Limonium*. *Annals of Botany* **17**: 615–627.
- 635 **Baker HG. 1955.** Self-compatibility and establishment after ‘long-distance’ dispersal.  
636 *Evolution* **9**: 347–348.
- 637 **Baker HG. 1959.** The contribution of autecological and genecological studies to our  
638 knowledge of the past migrations of plants. *American Naturalist* **93**: 255–272.
- 639 **Baker HG. 1966.** The evolution, functioning and breakdown of heteromorphic  
640 incompatibility systems, I. The Plumbaginaceae. *Evolution* **20**: 349–368.
- 641 **Baker HG. 1967.** Support for Baker’s Law - as a rule. *Evolution* **21**: 853–856.
- 642 **Barrett SCH. 1989.** The evolutionary breakdown of heterostyly. In: Bock JH, Linhart  
643 YB, eds. *The evolutionary ecology of plants*. Bolder, USA: Westview Press, 151–  
644 169.
- 645 **Barrett SCH. 1992.** Heterostylous genetic polymorphisms: model systems for  
646 evolutionary analysis. In: Barrett SCH, ed. *Evolution and function of heterostyly*.  
647 Berlin, Germany: Springer-Verlag, 1–29.

- 648 **Barrett SCH, Jesson LK, Baker AM. 2000.** The evolution and function of stylar  
649 polymorphisms in flowering plants. *Annals of Botany* **85**: 253–265.
- 650 **Barrett SCH, Shore JS. 2008.** New insights on heterostyly: comparative biology,  
651 ecology and genetics. In: Franklin-Tong VE, ed. *Self-incompatibility in flowering*  
652 *plants – evolution, diversity, and mechanisms*. Berlin, Germany: Springer-Verlag, 3–  
653 32.
- 654 **Bell CD, Soltis DE, Soltis PS. 2010.** The age and diversification of the angiosperms re-  
655 revisited. *American Journal of Botany* **97**: 1296–1303.
- 656 **Boissier E. 1848.** Plumbaginales. In: de Candolle AP, ed. *Prodromus systematis*  
657 *naturalis regni vegetabilis* Vol. 12. Paris, France: Treuttel et Wurz, 617–696.
- 658 **Bokhari MH. 1972.** A brief review of stigma and pollen types in *Acantholimon* and  
659 *Limonium*. *Notes of the Royal Botanic Garden of Edinburgh* **32**: 79–84.
- 660 **Bull JJ, Charnov LL. 1985.** On irreversible evolution. *Evolution* **39**: 1149–1155.
- 661 **Capella-Gutiérrez S, Silla-Martínez JM, Gabaldón T. 2009.** trimAl: a tool for  
662 automated alignment trimming in large-scale phylogenetic analyses. *Bioinformatics*  
663 **25**: 1972–1973.
- 664 **Caperta AD, Castro S, Loureiro J, Róis AS, Conceição S, Costa J, Rhazi L,**  
665 **Espírito-Santo D, Arsénio P. 2016.** Biogeographical, ecological and ploidy  
666 variation in related asexual and sexual *Limonium* taxa (Plumbaginaceae). *Botanical*  
667 *Journal of the Linnean Society*. doi: <https://doi.org/10.1111/boj.12498>
- 668 **Carman JG. 1997.** Asynchronous expression of duplicate genes in angiosperms may  
669 cause apomixis, bispory, tetraspory, and polyembryony. *Botanical Journal of the*  
670 *Linnean Society* **61**: 51–94.
- 671 **Cesaro AD, Thompson JD. 2004.** Darwin's cross-promotion hypothesis and the  
672 evolution of stylar polymorphism. *Ecology Letters* **7**: 1209–1215.
- 673 **Charlesworth D, Charlesworth B. 1979.** A model for the evolution of distyly.  
674 *American Naturalist* **114**: 467–498.
- 675 **Cohen JI. 2010.** “A case to which no parallel exists”: the influence of Darwin’s  
676 *Different Forms of Flowers*. *American Journal of Botany* **97**: 701–716.



- 677 **Cohen JI. 2013.** A phylogenetic analysis of morphological and molecular characters of  
678 Boraginaceae: evolutionary relationships, taxonomy, and patterns of character  
679 evolution. *Cladistics* **30**: 1–31.
- 680 **Costa J, Castro S, Loureiro J, Barrett SCH. 2017.** Experimental insights on the  
681 function of ancillary pollen and stigma polymorphisms in plants with heteromorphic  
682 incompatibility. *Evolution* **71**: 121–134.
- 683 **Cowan R, Ingrouille MJ, Lledó MD. 1998.** The taxonomic treatment of agamosperms  
684 in the genus *Limonium* Mill. (Plumbaginaceae). *Folia Geobotanica* **33**: 353–366.
- 685 **Crespo MB. 2009.** A new coastal species of *Limonium* (Plumbaginaceae) from  
686 Southeastern Spain. *Folia Geobotanica* **44**: 177–190.
- 687 **D'Amato F. 1940.** Contributo all'embriologia delle Plumbaginaceae. *Nuovo Giornale*  
688 *Botanico Italiano* **47**: 349–382.
- 689 **D'Amato F. 1949.** Triploidia e apomissia in *Statice oleifolia* Scop. var. *confusa* Godr.  
690 *Caryologia* **401**: 157–161.
- 691 **Dahlgren KVO. 1918.** Heterostylie innerhalb der Gattung *Plumbago*. *Svensk Botanisk*  
692 *Tidskrift* **12**: 362–372.
- 693 **Darwin C. 1877.** *The different forms of flowers on plants of the same species*. London:  
694 John Murray.
- 695 **Dollo L. 1893.** Les lois de l'évolution. *Bulletin de la Société Belge de Géologie,*  
696 *Paleontologie, et Hydrologie.* **7**: 164–166.
- 697 **Dulberger R. 1975a.** Intermorph structural differences between stigmatic papillae and  
698 pollen grains in relation to incompatibility in Plumbaginaceae. *Proceedings of the*  
699 *Royal Society of London B* **188**: 257–274.
- 700 **Dulberger R. 1975b.** S-gene action and the significance of characters in the  
701 heterostylous syndrome. *Heredity* **35**: 407–415.
- 702 **Dulberger R. 1992.** Floral polymorphisms and their functional significance in the  
703 heterostylous syndrome. In: Barrett SCH, ed. *Evolution and function of heterostyly*.  
704 Berlin, Germany: Springer-Verlag, 41–84.
- 705 **eFloras. 2008.** Published on the Internet <http://www.efloras.org> [accessed December



- 706 30, 2016] Missouri Botanical Garden, St. Louis, MO & Harvard University Herbaria,  
707 Cambridge, MA.
- 708 **Erben M. 1979.** Karyotype differentiation and its consequences in Mediterranean  
709 *Limonium*. *Webbia* **34**: 409–417.
- 710 **Erben M. 1993.** *Limonium*. In: Castroviejo S, Lainz M, González G, Montserrat P,  
711 Garmendia F, Paiva J, Villar L, eds. *Flora Iberica Vol. III*. Madrid, Spain: Real  
712 Jardín Botánico, C.S.I.C., 2–143.
- 713 **Erdtman G. 1986.** *Pollen morphology and plant taxonomy: Angiosperms*. Leiden, The  
714 Netherlands: E. J. Brill.
- 715 **Ferrero V, Arroyo J, Castro S, Navarro L. 2012.** Unusual heterostyly: style  
716 dimorphism and self-incompatibility are not tightly associated in *Lithodora* and  
717 *Glandora* (Boraginaceae). *Annals of Botany* **109**: 655–665.
- 718 **Ferrero V, Arroyo J, Vargas P, Thompson JD, Navarro L. 2009a.** Evolutionary  
719 transitions of style polymorphisms in *Lithodora* (Boraginaceae). *Perspectives in*  
720 *Plant Ecology, Evolution and Systematics* **11**: 111–125.
- 721 **Ferrero V, de Vega C, Stafford GI, Van Staden J, Johnson SD. 2009b.** Heterostyly  
722 and pollinators in *Plumbago auriculata* (Plumbaginaceae). *South African Journal of*  
723 *Botany* **75**: 778–784.
- 724 **Franklin-Tong VE (ed). 2008.** *Self-incompatibility in flowering plants - evolution,*  
725 *diversity, and mechanisms*. Berlin, Germany: Springer-Verlag.
- 726 **Ganders FR. 1979.** The biology of heterostyly. *New Zealand Journal of Botany* **17**:  
727 607–635.
- 728 **Ghobary MOW-E. 1984.** The systematic relationships of *Aegialitis* (Plumbaginaceae)  
729 as revealed by pollen morphology. *Plant Systematics and Evolution* **144**: 53–58.
- 730 **Ghobary MOW-E. 1986.** Dimorphic exine sculpturing in two distylous species of  
731 *Dyerophytum* (Plumbaginaceae). *Plant Systematics and Evolution* **152**: 267–276.
- 732 **Goldberg EE, Igic B. 2008.** On phylogenetic tests of irreversible evolution. *Evolution*  
733 **62**: 2727–2741.
- 734 **Graham SW, Barrett SCHB. 2004.** Phylogenetic reconstruction of the evolution of

- 735 stylar polymorphisms in *Narcissus* (Amaryllidaceae). *American Journal of Botany*  
736 **91**: 1007–1021.
- 737 **Heslop-Harrison J, Heslop-Harrison Y. 1985.** Surfaces and secretions in the pollen-  
738 stigma interaction: a brief review. *Journal of Cell Science. Supplement 2*: 287–300.
- 739 **Huelsbeck JP, Nielsen R, Bollback JP. 2003.** Stochastic mapping of morphological  
740 characters. *Systematic Biology* **52**: 131–158.
- 741 **Ingrouille MJ, Stace CA. 1985.** Pattern of variation of agamosperous *Limonium*  
742 (Plumbaginaceae) in the British Isles. *Nordic Journal of Botany* **5**: 113–125.
- 743 **Iversen J. 1940.** Blütenbiologische Studien. I. Dimorphie und monomorphie bei  
744 *Armeria*. *Kgl. Danske Videnskabernes Selskab. Biologiske Meddelelser*. **15**: 1–39.
- 745 **Katoh K, Standley DM. 2013.** MAFFT multiple sequence alignment software version  
746 7: Improvements in performance and usability. *Molecular Biology and Evolution* **30**:  
747 772–780.
- 748 **Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S,**  
749 **Cooper A, Markowitz S, Duran C, et al. 2012.** Geneious Basic: an integrated and  
750 extendable desktop software platform for the organization and analysis of sequence  
751 data. *Bioinformatics* **28**: 1647–1649.
- 752 **Keller B, Thomson JD, Conti E. 2014.** Heterostyly promotes disassortative pollination  
753 and reduces sexual interference in Darwin's primroses: evidence from experimental  
754 studies. *Functional Ecology* **28**: 1413–1425.
- 755 **Kissling J, Barrett SCH. 2013.** Variation and evolution of herkogamy in *Exochaenium*  
756 (Gentianaceae): Implications for the evolution of distyly. *Annals of Botany* **112**: 95–  
757 102.
- 758 **Kohn JR, Barrett SCH. 1992.** Experimental studies on the functional significance of  
759 heterostyly. *Evolution* **46**: 43–55.
- 760 **Kubitzki K. 1993.** Plumbaginaceae. In: Kubitzki K, Rohwer JG, Bittrich V, eds. *The*  
761 *families and genera of vascular plants*. Berlin, Germany: Springer-Verlag, 523–530.
- 762 **Kück P, Meusemann K. 2010.** FASconCAT. Forschungsmuseum A. Koenig,  
763 Germany.

- 764 **Lledó MD, Crespo MB, Cameron KM, Fay MF, Chase W. 1998.** Systematics of  
765 Plumbaginaceae based upon cladistic analysis of *rbcL* sequence data. *Systematic*  
766 *Botany* **23**: 21–29.
- 767 **Lledó MD, Crespo MB, Fay MF, Chase MW. 2005.** Molecular phylogenetics of  
768 *Limonium* and related genera (Plumbaginaceae): biogeographical and systematic  
769 implications. *American Journal of Botany* **92**: 1189–1198.
- 770 **Lledó MD, Karis PO, Crespo MB, Fay MF, Chase MW. 2001.** Phylogenetic position  
771 and taxonomic status of the genus *Aegialitis* and subfamilies Staticoideae and  
772 Plumbaginoideae (Plumbaginaceae): evidence from plastid DNA sequences and  
773 morphology. *Plant Systematics and Evolution* **229**: 107–124.
- 774 **Lloyd DG. 1992.** Self- and cross-fertilization in plants. II. The selection of self-  
775 fertilization. *International Journal of Plant Sciences* **153**: 370–380.
- 776 **Lloyd DG, Webb CJ. 1992a.** The evolution of heterostyly. In: Barrett SCH, ed.  
777 *Evolution and function of heterostyly*. Berlin, Germany: Springer-Verlag, 151–178.
- 778 **Lloyd DG, Webb CJ. 1992b.** The selection of heterostyly. In: Barrett SCH, ed.  
779 *Evolution and function of heterostyly*. Berlin, Germany: Springer-Verlag, 179–208.
- 780 **Maddison WP, Maddison DR. 2015.** Mesquite: a modular system for evolutionary  
781 analysis. Version 3.04. Available at <http://mesquiteproject.org>
- 782 **Mattsson O. 1983.** The significance of exine oils in the initial interaction between  
783 pollen and stigma in *Armeria maritima*. In: Mulchay DL, Ottaviano E, eds. *Pollen:*  
784 *biology and implications for plant breeding*. Elsevier Science Publishing Co., 257–  
785 264.
- 786 **McDill J, Replinger M, Simpson BB, Kadereit JW. 2009.** The phylogeny of *Linum*  
787 and Linaceae subfamily Linoideae, with implications for their systematics,  
788 biogeography, and evolution of heterostyly. *Systematic Botany* **34**: 386–405.
- 789 **Miller MA, Pfeiffer W, Schwartz T. 2010.** Creating the CIPRES Science Gateway for  
790 inference of large phylogenetic trees. *Proceedings of the Gateway Computing*  
791 *Environments Workshop (GCE)*. New Orleans, 1–8.
- 792 **Morretti F, Puppi G, Giuliani C, Conti F. 2015.** Heterostyly in *Goniolimon italicum*  
793 (Plumbaginaceae), endemic to Abruzzo (central Apennines, Italy). *Anales del Jardín*

- 794 *Botánico de Madrid* **72**: 1–5.
- 795 **Pagel M, Meade A, Barker D. 2004.** Bayesian estimation of ancestral character states  
796 on phylogenies. *Systematic Biology* **53**: 673–684.
- 797 **Pannell JR, Auld JR, Brandvain Y, Burd M, Busch JW, Cheptou P, Conner JK,**  
798 **Goldberg EE, Grant A, Grossenbacher DL, et al. 2015.** The scope of Baker 's  
799 law. *New Phytologist* **208**: 656–667.
- 800 **Paradis E. 2013.** Molecular dating of phylogenies by likelihood methods: a comparison  
801 of models and a new information criterion. *Molecular Phylogenetics and Evolution*  
802 **67**: 436–444.
- 803 **Paradis E, Claude J, Strimmer K. 2004.** APE: analysis of phylogenetics and  
804 evolution in R language. *Bioinformatics* **20**: 289–290.
- 805 **de Queiroz A, Gatesy J. 2007.** The supermatrix approach to systematics. *Trends in*  
806 *Ecology & Evolution* **22**: 34–41.
- 807 **Revell LJ. 2012.** phytools: an R package for phylogenetic comparative biology (and  
808 other things). *Methods in Ecology and Evolution* **3**: 217–223.
- 809 **Róis AS, Sádio F, Paulo OS, Teixeira G, Paes AP, Espírito-Santo D, Sharbel TF,**  
810 **Caperta AD. 2016.** Phylogeography and modes of reproduction in diploid and  
811 tetraploid halophytes of *Limonium* species (Plumbaginaceae): evidence for a pattern  
812 of geographical parthenogenesis. *Annals of Botany* **117**: 37–50.
- 813 **Roquet C, Thuiller W, Lavergne S. 2013.** Building megaphylogenies for  
814 macroecology: taking up the challenge. *Ecography* **36**: 13–26.
- 815 **Roskov Y, Abucay L, Orrell T, Nicolson D, Flann C, Bailly N, Kirk P, Bourgoi T,**  
816 **DeWalt RE, Decock W, et al.** Species 2000 & ITIS Catalogue of Life, 2016 Annual  
817 Checklist. Digital resource at [www.catalogueoflife.org/annual-checklist/2016](http://www.catalogueoflife.org/annual-checklist/2016)  
818 Species 2000: Naturalis, Leiden, the Netherlands. ISSN 2405-884X.
- 819 **Sánchez R, Serra F, Tárraga J, Medina I, Carbonell J, Pulido L, De María A,**  
820 **Capella-Gutiérrez S, Huerta-Cepas J, Gabaldón T, et al. 2011.** Phylemon 2.0: a  
821 suite of web-tools for molecular evolution, phylogenetics, phylogenomics and  
822 hypotheses testing. *Nucleic Acids Research* **39**: 1–5.

- 823 **Schill R, Baumm A, Wolter M. 1985.** Comparative micromorphology of stigma  
824 surfaces in angiosperms; relations to pollen grain surfaces in heterostylous taxa.  
825 *Plant Systematics and Evolution* **148**: 185–214.
- 826 **Soltis DE, Soltis PS, Chase MW, Mort ME, Albach DC, Zanis M, Savolainen V,**  
827 **Hahn WH, Hoot SB, Fay MF, et al. 2000.** Angiosperm phylogeny inferred from  
828 18S rDNA, *rbcL*, and *atpB* sequences. *Botanical Journal of the Linnean Society*  
829 **133**: 381–461.
- 830 **Souza LGR, Crosa O, Speranza P, Guerra M. 2012.** Cytogenetic and molecular  
831 evidence suggest multiple origins and geographical parthenogenesis in  
832 *Nothoscordum gracile* (Alliaceae). *Annals of Botany* **109**: 987–999.
- 833 **Stamatakis A. 2014.** RAxML Version 8: a tool for phylogenetic analysis and post-  
834 analysis of large phylogenies. *Bioinformatics*. doi 10.1093/bioinformatics/btu033.
- 835 **Stebbins GL. 1974.** *Flowering plants: evolution above the species level*. Cambridge:  
836 Harvard University Press.
- 837 **Stone JL, Thomson JD. 1994.** The evolution of distyly: pollen transfer in artificial  
838 flowers. *Evolution* **48**: 1595-1606.
- 839 **Stöver BC, Müller KF. 2010.** TreeGraph 2: combining and visualizing evidence from  
840 different phylogenetic analyses. *BMC Bioinformatics* **11**: 7.
- 841 **The Plant List. 2013.** The Plant List - Version 1.1. Available online  
842 <http://www.theplantlist.org/>
- 843 **Tippery NP, Les DH. 2011.** Phylogenetic relationships and morphological evolution in  
844 *Nymphoides* (Menyanthaceae). *Systematic Botany* **36**: 1101–1113.
- 845 **Torices R, Anderberg AA. 2009.** Phylogenetic analysis of sexual systems in Inuleae  
846 (Asteraceae). *American Journal of Botany* **96**: 1011–1019.
- 847 **Webb CJ, Lloyd DG. 1986.** The avoidance of interference between the presentation of  
848 pollen and stigmas in angiosperms II. Herkogamy. *New Zealand Journal of Botany*  
849 **24**: 163-178.
- 850 **Weller SG. 2009.** The different forms of flowers - what have we learned since Darwin?  
851 *Botanical Journal of the Linnean Society* **160**: 249-261.

852 **Zhou W, Barrett SCH, Wang H, Li D-Z. 2015.** Reciprocal herkogamy promotes  
 853 disassortative mating in a distylous species with intramorph compatibility. *New*  
 854 *Phytologist* **206**: 1503-1512.

855

856

## 857 **Figure captions**

858

859 **Figure 1** Phylogenetic distribution of the six characters investigated in Plumbaginaceae.  
 860 Subfamily Plumbaginoideae is represented by the clade containing *Plumbago*, whereas  
 861 subfamily Staticoideae is represented by the clades containing the genera *Acantholimon*,  
 862 *Goniolimon*, *Armeria*, *Psylliostachys* and *Limonium*. **(a)** Reciprocal herkogamy: black –  
 863 reciprocal herkogamy, grey – style-stamen height monomorphism. **(b)** Incompatibility  
 864 system: blue – self-incompatible, yellow – self-compatible. **(c)** Exine sculpturing: green  
 865 – monomorphic, red - *Armeria*-type pollen. **(d)** Stigmatic papillae condition: dark blue –  
 866 dimorphic, light blue – monomorphic. **(e)** Stigma morphology: brown – capitate, light  
 867 pink – filiform, dark pink – others (i.e., fimbriate, pear-shaped, lanceolate). **(f)**  
 868 Reproductive mode: yellow – sexual, orange – apomictic. In all cases, white represents  
 869 missing information for a given species. Pie diagrams show the posterior probability of  
 870 reciprocal herkogamy (black) vs style-stamen height monomorphism (grey), self-  
 871 incompatibility (blue) vs self-compatibility (yellow), monomorphic (green) vs *Armeria*-  
 872 type pollen (red), and dimorphic (dark blue) vs monomorphic (light) stigmas for a given  
 873 node.

874

875 **Figure 2** Evolutionary buildup and breakdown of heterostyly in Plumbaginaceae: **(c)**-  
 876 **(d)** subfamily Plumbaginoideae, **(e)-(g)** subfamily Staticoideae. Steps (a)-(c) correspond  
 877 to the selfing avoidance model of Charlesworth & Charlesworth (1979); stigma-height  
 878 dimorphism is omitted as there is no evidence of this intermediate stage in the family.  
 879 Hypothesized selective forces are indicated in italics next to solid arrows, which  
 880 indicate transitions between states. Dashed arrows in (c) and (g) indicate compatible  
 881 pollinations. **(a)** Self-compatible (SC) ancestor with stigmas and anthers at the same  
 882 height. **(b)** Evolution of heteromorphic incompatibility with two mating groups with  
 883 monomorphic sex organs, pollen and stigmas. **(c)** Evolution of distyly, heteromorphic  
 884 incompatibility and monomorphic pollen and stigmas in *Plumbago*, *Dyerophytum* and



885 *Cerastostigma* species. **(d)** Evolution of homostyles, SC with monomorphic pollen and  
 886 stigmas and smaller-flowered *Plumbagella micrantha*. **(e)** Evolution of pollen-stigma  
 887 dimorphism in *Acantholimon*, *Goniolimon*, *Armeria*, *Psylliostachys*, *Ceratolimon*,  
 888 *Limoniastrum* and *Limonium* species. Top line, left to right: pollen type A, pollen type  
 889 B. Bottom line, left to right: cob stigmas, papillate stigmas. Pollen and stigma images  
 890 from Figs. 1-2 in Costa *et al.* (2017). Transition to homostyly in Statioideae is based  
 891 on Baker (1948c). **(f)** Apomictic small-flowered species of *Limonium* possessing A/cob  
 892 and B/papillate pollen-stigma combinations. **(g)** Distylous, SI and pollen-stigma  
 893 dimorphism in *Limonium vulgare*, *Acantholimon* and *Goniolimon* species. Downwards  
 894 arrow from (g) to (e) indicates the breakdown of distyly in *Acantholimon* species.

895

896 **Figure 3** Transition rates between the four character-state combinations of reproductive  
 897 mode and pollen-stigma dimorphism for species of Plumbaginaceae in the phylogeny ( $n$   
 898 = 121 species). For seven species, there was no information regarding character state-  
 899 combinations considered.  $q_{ij}$  – transition from state  $i$  to state  $j$ ; reproductive system: Sex  
 900 – sexual, A – apomictic; pollen-stigma dimorphism: M – monomorphic pollen and  
 901 stigmas, PSD – pollen-stigma dimorphism;  $n$  – number of taxa in each character state  
 902 combination.

903

904

### 905 **Supporting Information**

906 **Fig. S1** Ultrametric maximum likelihood phylogenetic tree of Plumbaginaceae.

907 **Methods S1** Characterization of the floral polymorphism and associated characters.

908 **Table S1** Information on each herbarium specimen sampled for data on reciprocal  
 909 herkogamy and ancillary characters.

910 **Table S2** Character states used in the ancestral character state reconstruction of  
 911 Plumbaginaceae.

912 **Table S3** GenBank accession number for all taxa included in the supermatrix for  
 913 maximum likelihood analysis of Plumbaginaceae.

914 **Table S4** Models of substitution rate variation among branches and values of the  
 915 smoothing parameter, lambda, tested for creating an ultrametric tree of Plumbaginaceae.



916 **Table S5** Support for models of character trait evolution for the most recent common  
917 ancestor of the Plumbaginaceae following Bayesian analyses (BIC) and stochastic  
918 character mapping (PP) for the second set of trees.

919 **Table S6** Rate of change between states for the six characters investigated for the  
920 second set of 100 trees of Plumbaginaceae.

921 **Table S7** Support for models of correlated evolution between traits in Plumbaginaceae  
922 and the values of transition rates ( $q_{ij}$ ) obtained from the models.

For Peer Review

923 **Table 1** Support for models of character state evolution for the most recent common  
 924 ancestor of the Plumbaginaceae following Bayesian Markov chain Monte Carlo  
 925 analyses (BIC) and stochastic character mapping (PP).

<b>Model</b>	<b>BIC</b>	<b>PP</b>
(a) Sex-organ arrangement, ER		
Monomorphism	67.622	0.966
Reciprocal herkogamy	72.706	0.034
(b) Incompatibility status, ARD		
Self-incompatible	50.052	1.000
Self-compatible	55.480	0.000
(c) Exine sculpturing, ER		
Monomorphic	106.564	0.773
Dimorphic (i.e., <i>Armeria</i> -type)	108.670	0.227
(d) Stigmatic papillae shape, ER		
Monomorphic	99.594	0.570
Dimorphic	100.444	0.430
(e) Reproductive mode, ARD		
Sexual	45.518	1.000
Apomictic	51.102	0.000
(f) Stigma morphology, ER		
Filiform	67.382	0.091
Capitate	63.754	0.881
Others	69.968	0.027

926 ER – Equal rates model, ARD – All rates different model, BIC – Bayesian Information Criterion, PP –  
 927 Posterior Probability.

928

929

930

931

932 **Table 2** Number, frequency and rate of change between states for the six characters  
 933 investigated for the first set of 100 trees of Plumbaginaceae.

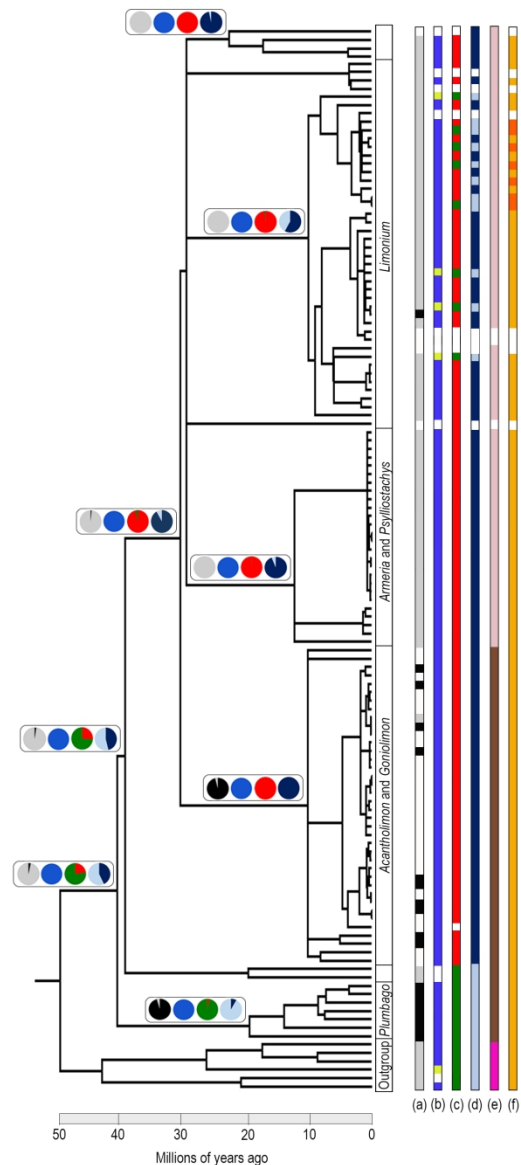
Character state	No. of transitions	Frequency of transition	Relative rate of transition
(a) Sex-organ arrangement			
Monomorphic to RH	3.70 ± 0.01	0.57	0.75
RH to homostylous	2.83 ± 0.01	0.43	1.80
(b) Incompatibility status			
SI to SC	4.99 ± 0.003	0.99	1.02
SC to SI	0.05 ± 0.01	0.01	0.35
(c) Exine sculpturing			
<i>Armeria</i> -type to monomorphic	9.28 ± 0.02	0.80	1.42
Monomorphic to <i>Armeria</i> -type	2.39 ± 0.01	0.20	0.47
(d) Stigmatic papillae shape			
Dimorphic to monomorphic	11.08 ± 0.03	0.59	1.06
Monomorphic to dimorphic	7.55 ± 0.03	0.41	0.92
(e) Reproductive mode			
Apomictic to sexual	0.29 ± 0.03	0.05	0.95
Sexual to apomictic	5.01 ± 0.02	0.95	6.88
(f) Stigma morphology			
Filiform to capitate	0.86 ± 0.03	0.14	0.33
Filiform to others	0.58 ± 0.03	0.10	0.22
Capitate to filiform	1.37 ± 0.03	0.23	0.54
Capitate to others	1.31 ± 0.04	0.22	0.51
Others to filiform	0.75 ± 0.02	0.13	1.01
Others to capitate	1.08 ± 0.03	0.18	1.45

934 Values are mean ± SE. RH – reciprocal herkogamy, SI – self-incompatible, SC – self-compatible. For  
 935 details of the calculation of frequency of transitions and relative rates of transition see *Material and*  
 936 *Methods*.

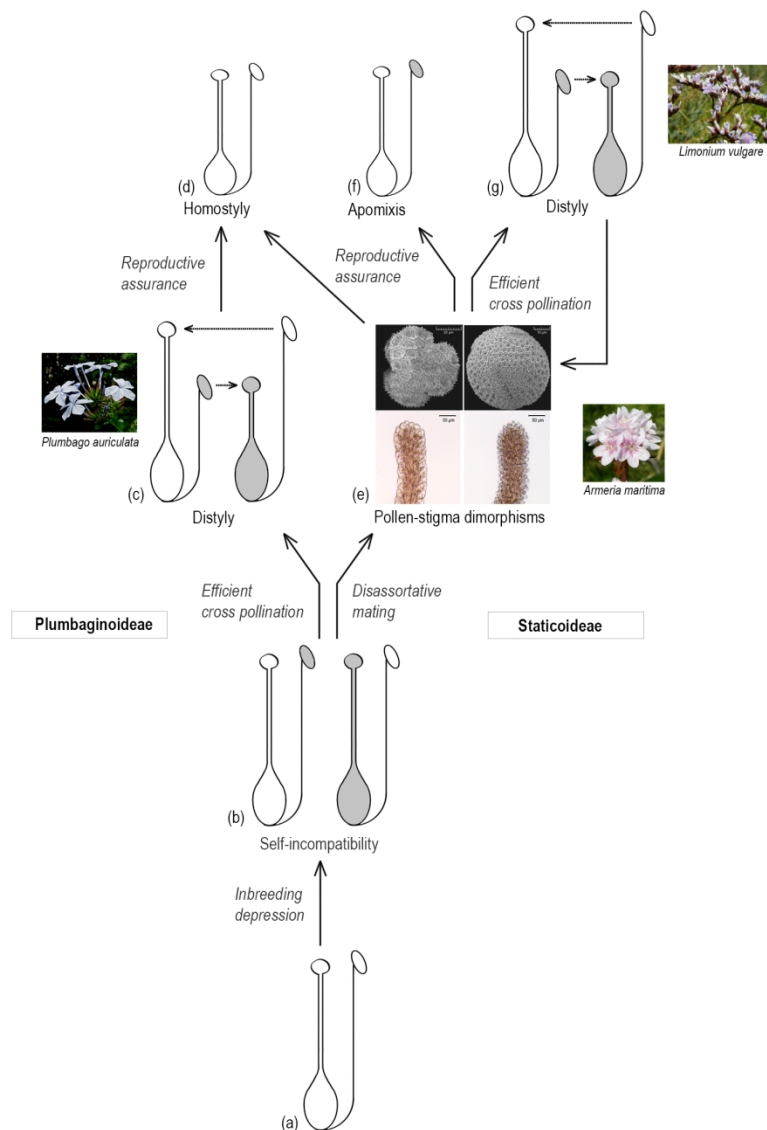
937

938

939



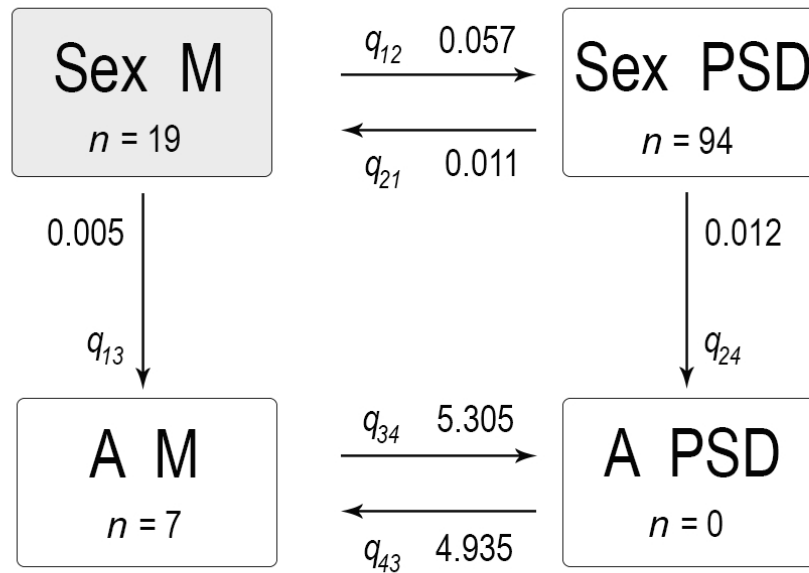
**Figure 1** Phylogenetic distribution of the six characters investigated in Plumbaginaceae. Subfamily Plumbaginoideae is represented by the clade containing *Plumbago*, whereas subfamily Statioideae is represented by the clades containing the genera *Acantholimon*, *Goniolimon*, *Armeria*, *Psylliostachys* and *Limonium*. **(a)** Reciprocal herkogamy: black – reciprocal herkogamy, grey – style-stamen height monomorphism. **(b)** Incompatibility system: blue – self-incompatible, yellow – self-compatible. **(c)** Exine sculpturing: green – monomorphic, red – *Armeria*-type pollen. **(d)** Stigmatic papillae condition: dark blue – dimorphic, light blue – monomorphic. **(e)** Stigma morphology: brown – capitate, light pink – filiform, dark pink – others (i.e., fimbriate, pear-shaped, lanceolate). **(f)** Reproductive mode: yellow – sexual, orange – apomictic. In all cases, white represents missing information for a given species. Pie diagrams show the posterior probability of reciprocal herkogamy (black) vs style-stamen height monomorphism (grey), self-incompatibility (blue) vs self-compatible (yellow), monomorphic (green) vs *Armeria*-type pollen (red), and dimorphic (dark blue) vs monomorphic (light) stigmas for a given node.



**Figure 2** Evolutionary buildup and breakdown of heterostyly in Plumbaginaceae: **(c)-(d)** subfamily Plumbaginoideae, **(e)-(g)** subfamily Staticoideae. Steps (a)-(c) correspond to the selfing avoidance model of Charlesworth & Charlesworth (1979); stigma-height dimorphism is omitted as there is no evidence of this intermediate stage in the family. Hypothesized selective forces are indicated in italics next to solid arrows, which indicate transitions between states. Dashed arrows in (c) and (g) indicate compatible pollinations. **(a)** Self-compatible (SC) ancestor with stigmas and anthers at the same height. **(b)** Evolution of heteromorphic incompatibility with two mating groups with monomorphic sex organs, pollen and stigmas. **(c)** Evolution of distyly, heteromorphic incompatibility and monomorphic pollen and stigmas in *Plumbago*, *Dyerophytum* and *Cerastostigma* species. **(d)** Evolution of homostyles, SC with monomorphic pollen and stigmas and smaller-flowered *Plumbagella micrantha*. **(e)** Evolution of pollen-stigma dimorphism in *Acantholimon*, *Goniolimon*, *Armeria*, *Psylliostachys*, *Ceratolimon*, *Limoniasstrum* and *Limonium* species. Top line, left to right: pollen type A, pollen type B. Bottom line, left to right: cob stigmas, papillate stigmas. Pollen and stigma images from Figs. 1-2 in Costa *et al.* (2017). Transition to homostyly in Staticoideae is based on Baker (1948c). **(f)** Apomictic small-flowered species of *Limonium* possessing A/cob and B/papillate pollen-stigma combinations.

**(g)** Distylous, SI and pollen-stigma dimorphism in *Limonium vulgare*, *Acantholimon* and *Goniolimon* species. Downwards arrow from (g) to (e) indicates the breakdown of distyly in *Acantholimon* species.





**Figure 3** Transition rates between the four character-state combinations of reproductive mode and pollen-stigma dimorphism for species of Plumbaginaceae in the phylogeny ( $n = 121$  species). For seven species, there was no information regarding character state-combinations considered.  $q_{ij}$  – transition from state  $i$  to state  $j$ ; reproductive system: Sex – sexual, A – apomictic; pollen-stigma dimorphism: M – monomorphic pollen and stigmas, PSD – pollen-stigma dimorphism;  $n$  – number of taxa in each character state combination.